

***In vitro* antimicrobial activity of tigecycline against Gram negative and Gram positive pathogens collected in Northern Italy (T.E.S.T. program 2010)**

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SUMMARY

Background. In this study (part of the global T.E.S.T. program) was evaluated the *in vitro* activity of tigecycline, member of a new class of antimicrobial agents, the glycylycyclines, against clinical isolates collected in Italy.

Methods. A total of 194 clinical isolates were collected and identified in our Institution during 2010. Minimum inhibitory concentrations (MICs) of the antimicrobial agents were determined by the CLSI (2010) recommended broth microdilution method.

Results. Globally 129 Gram negative and 65 Gram positive pathogens were evaluated. Tigecycline demonstrated excellent inhibitory activity against *Escherichia coli*, *Haemophilus influenzae*, *Enterococcus* spp., *Staphylococcus aureus* MetS, *Streptococcus pneumoniae* and *Streptococcus agalactiae* with MIC₉₀ ≤ 1 mg/l.

Conclusion. Tigecycline exhibited potent *in vitro* antibacterial activity (comparable to or greater than most commonly employed antimicrobials) against both Gram positive and negative clinical pathogens. These data suggest that tigecycline, with an expanded broad-spectrum antimicrobial activity, may be an effective empiric therapeutic option for the treatment of serious infections caused by clinically relevant pathogens.

INTRODUCTION

Tigecycline is a first-in-class of expanded broad-spectrum glycylycycline. It inhibits bacterial protein synthesis by binding to the 30s ribosomal subunit, but with five times higher affinity than tetracyclines (2). This new drug was approved for use in Europe in 2006 for complicated skin and soft-tissue and intra-abdominal infections (<http://www.emea.europa.eu/humandocs/Humans/EPAR/tygacil/tygacil.htm>).

In vitro studies demonstrate that it has good activity against many commonly encountered respiratory bacteria, including multiple resistant Gram positive, Gram negative, anaerobic, as well as multidrug-resistant (MDR) pathogens such as methicillin-resistant *S. aureus* (MRSA) and *S. epidermidis* (MRSE), vancomycin-resistant *Enterococcus* spp. (VRE), penicillin-resistant *S. pneumoniae* (PRSP) and β -lactamase producing *H. influenzae* (3).

Tigecycline inhibits bacterial protein biosynthesis blocking the attachment of amino-acyl tRNA to the A site of the ribosome and preventing the elongation of peptide chains (6).

The drug maintains its activity even in presence of efflux pumps (encoded by *tetA-tetD* and *tetK* genes) and ribosomal protection (*tetM*) mechanisms that otherwise confer tetracycline resistance. Tigecycline appears to overcome these mechanisms of resistance because of steric hin-

drance due to the addition of a large substituent on the D ring at the 9th position of the tetracycline molecule (7, 13).

This study is part of the larger global Tigecycline Evaluation and Surveillance Trials (T.E.S.T.) program and was undertaken to document the *in vitro* activity of tigecycline against clinical isolates collected in Northern Italy from in-patient and out-patient populations.

MATERIALS AND METHODS

A total of 194 clinical isolates were collected and identified to the species level in our Institution during 2010. Isolates were collected from both inpatients and outpatients with a documented infection in which the isolate collected was identified as the probable causative organism according to institutional criteria. Only one isolate per patient was accepted into the study.

Minimum inhibitory concentrations (MICs) of the antimicrobial agents were determined by the Clinical and Laboratory Standards Institute (CLSI) (11) recommended broth microdilution testing method. Overnight cultures of bacteria were diluted to give a final concentration of approximately 5×10^5 cells/ml. Samples were then added to equivalent volumes of the various concentration of antibiotics distributed on a microplate and prepared from serial twofold dilutions. After 18-24

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hours of incubation at 37°C, the concentrations of drugs that prevented visible growth were recorded as the MICs. Tigecycline was supplied by Wyeth Pharmaceuticals (Collegeville, PA, USA). All other agents were supplied by the panel manufacturer, MicroScan (Dade Behring Inc., Sacramento, CA, USA).

The panel of agents tested against Gram positive organisms was: tigecycline (0.008-16), amoxicillin/clavulanic acid (0.03-8), ampicillin (0.06-16), ceftriaxone (0.03-64), meropenem (0.12-16), linezolid (0.5-8), levofloxacin (0.06-32), minocycline (0.25-8), penicillin (0.06-8), piperacillin/tazobactam (0.25-16) and vancomycin (0.12-32). The panel of antimicrobial agents tested against Gram negative organisms was: tigecycline (0.008-16), amikacin (0.5-64), amoxicillin/clavulanic acid (0.12-32), ampicillin (0.5-32), cefepime (0.5-32), ceftazidime (8-32), ceftriaxone (0.06-64), levofloxacin (0.06-32) meropenem (0.12-16), minocycline (0.25-8) piperacillin/tazobactam (0.06-128). MIC interpretive criteria established by CLSI (11) and recent US Food and Drug Administration guidelines for tigecycline (Tygacil, Product Insert. Wyeth Pharmaceuticals, Inc., Philadelphia, PA, USA, 2005) were followed. As no interpretative criterion for tigecycline with *Acinetobacter baumannii* has been established, Clinical and laboratory Standards Institute interpretative criteria for *Enterobacteriaceae* were used for *A. baumannii*, as had been reported in previous literature (1, 17). Quality controls (QC) were performed using the

following strains: *Escherichia coli* ATCC 25922; *E. coli* ATCC 35218; *H. influenzae* ATCC 49766; *H. influenzae* ATCC 49247; *S. aureus* ATCC 29213; *P. aeruginosa* ATCC 27853; *Enterococcus faecalis* ATCC 29212 and *S. pneumoniae* ATCC 49619. Results were included in the analysis only when corresponding QC isolates tested within the acceptable range according to CLSI (2010).

RESULTS

The largest number of isolates was collected from in-patients population (medicine, surgery, ICU, nursing home\rehabilitation, renal, infectious diseases, paediatrics and obstetrics), only 35% of strains derived from community acquired infections (Table 1).

Globally 129 Gram negative and 65 Gram positive pathogens were evaluated. The first group of bacteria included 25 *E. coli*, 25 *Klebsiella* spp., 25 *Enterobacter* spp., 10 *Serratia marcescens*, 20 *P. aeruginosa*, 15 *A. baumannii*, (80% multi-drug resistant to cephalosporins aminoglycosides and quinolones simultaneously), 9 *H. influenzae* (11.1% β -lactamase-producers). Gram positive strains included 15 *Enterococcus* spp. (6.6% vancomycin-resistant), 25 *S. aureus* (68% methicillin-resistant), 15 *S. pneumoniae* (33.3% penicillin-non-susceptible), 10 *S. agalactiae*.

Bacterial isolates were obtained from blood (65, 33.5%), urine (17, 8.8%), respiratory tract (53, 27.3%), skin (20, 10.3%), wound (13, 6.7%), vagina swabs (13, 6.7%) and other site (8, 4.2%).

Table 1. Distribution of the strains collected in this study according to different clinical settings.

Microorganism	Tot	Community acquired	Hospital acquired									
			Med	Sur	ICU	Nursing home\ rehab	Ren	ID	Ped	Obs	Unk	Other
Gram negative												
<i>E. coli</i>	25	6	6	3	4	2	1					3
<i>Klebsiella</i> spp. (1)	25	17	1	6		1						
<i>Enterobacter</i> spp. (2)	25	4	5	6	4	1	1					4
<i>S. marcescens</i>	10	1			5						1	3
<i>P. aeruginosa</i>	20	10	1		5	1					1	2
<i>A. baumannii</i>	15	5	3		3	1						3
<i>H. influenzae</i>	9	2						2	3		1	1
Gram positive												
<i>S. agalactiae</i>	10	6				1				3		
<i>Enterococcus</i> spp. (3)	15	4		1	4	1	1		3			1
<i>S. aureus</i>	25	9	4	1	2	3	1				1	4
<i>S. pneumoniae</i>	15	4			2				8		1	

Med: medicine, Sur: surgery, ICU: Intensive Care Unit, NH\REH: Nursing home\ rehabilitation, Ren: renal, ID: Infectious Disease, Ped: pediatrics, Obs: obstetrics, Unk: non-specified unit.

(1) *K. pneumoniae* 16, *K. oxytoca* 7; (2) *E. cloacae* 21, *E. aerogenes* 4; (3) *E. faecalis* 10, *E. faecium* 5.

(Table 2).

Tigecycline demonstrate an excellent inhibitory activity against *Enterobacteriaceae* (Table 3), indeed tigecycline's MIC₉₀ was minor or equal to 4 mg/l. Against *E. coli* and *Klebsiella* spp. only meropenem behaves better than tigecycline (MIC₉₀ 0.12mg/l and 0.25 mg/l respectively).

This new agent, as the other tetracyclines, showed limited *in vitro* activity against *P. aeruginosa* with MIC₉₀ values equal to 16mg/l (Table 4).

Tigecycline demonstrated a good inhibitory activity against *A. baumannii* (MIC₅₀ 1mg/l and MIC₉₀ 2mg/l) even on multi-drug resistant strain, with the exception of a single isolate which showed a MIC value corresponding to 8mg/l. Such result is consistent with recent reports of emergence of tigecycline-resistant *A. baumannii* after tigecycline therapy (1, 14) (Table 4). *In vitro* tigecycline's activity on *A. baumannii* was superior to beta-lactams, beta-lactams/beta-lactamase inhibitor combinations, cephalosporins, aminoglycosides and levofloxacin.

Tigecycline demonstrate excellent inhibitory activity against *H. influenzae* (MIC₅₀ and MIC₉₀ 0.25mg/l), included β-lactamase-producers strains (Table 4).

Against Gram positive isolates (Table 5), tigecycline shown an excellent activity against *Enterococcus* spp. The drug presented MIC₉₀ of 0.25 mg/l, regardless of susceptibility to vancomycin. This value was the lowest of all comparative agents, in particular was several folds lower than linezolid, minocycline and levofloxacin.

The drug has shown a potent inhibitory activity against *S. aureus* regardless of methicillin-resistant phenotype. Tigecycline, with MIC₅₀ and MIC₉₀ values of 0.25 and 0.5 mg/l, respectively, demonstrated *in vitro* activity comparable to vancomycin and greater than linezolid and levofloxacin against MRSA. In MSSA the MIC₅₀ and MIC₉₀ value was 0.25 mg/l and 0.5 mg/l respectively.

Tigecycline demonstrated potent inhibitory activity against *S. agalactiae* and *S. pneumoniae* (Table 6), even in penicillin-resistant strains, with MIC₉₀ values ≤ 1mg/l.

DISCUSSION

Resistance to currently available antibiotics and incidence of infection due to multidrug-resistant

Table 2. Distribution of the strains collected in this study according to the type of clinical samples.

Microorganisms	Blood	Urine	Respiratory tract	Skin	Wound	Vaginal swabs	Other
Gram negative							
<i>E. coli</i>	11	3	7			3	1
<i>Klebsiella</i> spp. (1)	9	3	9			4	
<i>Enterobacter</i> spp. (2)	12	2		5	6		
<i>S. marcescens</i>	7		3				
<i>P. aeruginosa</i>	4	1	8	4	2		1
<i>A. baumannii</i>		4	4	3	1		3
<i>H. influenzae</i>			9				
Gram positive							
<i>S. agalactiae</i>		1		2		7	
<i>Enterococcus</i> spp. (3)	12	2		1			
<i>S. aureus</i>	7	1	4	5	4	1	3
<i>S. pneumoniae</i>	3		12				
Total	65	17	56	20	13	15	8

bacteria has dramatically increased worldwide during the last twenty years. In presence of a serious infection, appropriate empirical antibiotic therapy can be life-saving and the choice of an antimicrobial to which the pathogens are susceptible may be critical. For these reasons, the development of new antimicrobial agents with expanded antibacterial spectrum (increased activity against Gram negative as well as Gram positive microorganisms) is more urgent than ever (5, 18).

Tigecycline's *in vitro* activity was comparable to or greater than most commonly prescribed antimicrobial against important clinical pathogens.

Tigecycline, for example, demonstrated on methicillin-resistant *S. aureus* (MRSA) *in vitro* activity comparable to vancomycin, the antimicrobial agents currently used for the treatment of serious staphylococcal infections and exhibits greater activity to linezolid against vancomycin-resistant *Enterococcus faecalis* and *Enterococcus faecium* (VRE). This promising compound may be useful even in the treatment of serious infections caused by resistant Gram negative strains with limited therapeutic options. Tigecycline has shown to be highly effective against *E. coli* and *Klebsiella* spp. *A. baumannii* is a problematic pathogen, particularly in ICUs. The results from TEST reveal that no antimicrobial agents tested was active against *A. baumannii*. Tigecycline was the only agent that shown a good inhibitory activity against *A. baumannii* that are commonly associated with serious nosocomial infections. Resistance of *Acinetobacter* to cephalosporins, aminoglycosides and quinolones is widespread with an

increasing of multi-drug resistance (9, 8). Tigecycline is a potent antimicrobial agent even against the infections due to this important pathogen.

The drug demonstrated a limited activity, similar to other tetracyclines, only against *P. aeruginosa*.

The *in vitro* activity of tigecycline observed in this study suggests that this drug is a suitable antimicrobial agent for empiric treatment of serious infections sustained by some of the commonly encountered pathogens.

Tigecycline is available as parenteral agent, has linear pharmacokinetics, long terminal half-life and is extensively distributed in tissues (15).

Some interesting applications for this drug may be in surgical wound infections (particularly following abdominal surgery), and as alternative therapeutic agent in patient with serious allergies to β -lactam antibiotics (10).

Results from Phase III clinical studies in the treatment of complicated skin and skin structure infection (cSSSI) and complicated intra-abdominal infection (cIAI) have demonstrated the potential of tigecycline used as monotherapy for the treatment of this infections: the drug showed equivalence to imipenem in cIAI and to vancomycin plus aztreonam in cSSSI (12, 16, 4, 19).

The *in vitro* and *in vivo* studies points out that, tigecycline, with an expanded broad-spectrum antimicrobial activity against Gram positive and negative bacteria, offers a new alternative for the treatment of infections caused by clinically relevant pathogens in which the emergence of resistance to previously active antibiotics has created to the physicians limitations in therapeutic options.

Table 3. In vitro activity of tigecycline and comparative antimicrobial agents against Enterobacteriaceae clinical pathogens.

Microorganisms	Drugs	MIC 50	MIC 90	MIC Range
				$\mu\text{g/L}$
<i>E. coli</i>	Tigecycline	0.5	1	0.06 - 8
	Amikacin	8	16	1 - ≥ 64
	Amoxicillin Clavulanic Acid	16	32	4 - ≥ 32
	Ampicillin	≥ 32	≥ 32	1 - ≥ 32
	Cefepime	≤ 0.5	≥ 32	4 - ≥ 32
	Ceftazidime	≤ 8	≥ 32	≤ 8 - ≥ 32
	Ceftriaxone	0.25	≥ 64	≤ 0.06 - ≥ 64
	Levofloxacin	8	≥ 8	0.015 - ≥ 8
	Meropenem	≤ 0.06	0.12	≤ 0.06 - 0.5
	Minocycline	8	≥ 16	1 ≥ 16
<i>Klebsiella spp.</i>	Piperacillin Tazobactam	≥ 128	≥ 128	0.06 - ≥ 128
	Tigecycline	0.5	2	0.25 - 2
	Amikacin	2	32	1 - 32
	Amoxicillin Clavulanic Acid	2	32	1 - ≥ 32
	Ampicillin	≥ 32	≥ 32	≥ 32
	Cefepime	≤ 0.05	≥ 32	≤ 0.05 - ≥ 32
	Ceftazidime	≤ 8	≥ 32	≤ 8 - ≥ 32
	Ceftriaxone	≤ 0.06	64	≤ 0.06 - ≥ 64
	Levofloxacin	0.12	≥ 8	0.03 - ≥ 8
	Meropenem	≤ 0.06	0.25	≤ 0.06 - 1
<i>Enterobacter spp.</i>	Minocycline	4	16	1 - ≥ 16
	Piperacillin Tazobactam	2	64	1 - ≥ 128
	Tigecycline	1	4	0.5 - ≥ 16
	Amikacin	2	64	1 - ≥ 64
	Amoxicillin Clavulanic Acid	≥ 32	≥ 32	32 - ≥ 32
	Ampicillin	≥ 32	≥ 32	16 - ≥ 32
	Cefepime	≤ 0.5	≥ 32	≤ 0.5 - ≥ 32
	Ceftazidime	≤ 8	≥ 32	≤ 8 - ≥ 32
	Ceftriaxone	2	≥ 64	0.12 - ≥ 32
	Levofloxacin	0.12	8	0.03 - ≥ 8
<i>S. marcescens</i>	Meropenem	0.25	8	≤ 0.06 - 8
	Minocycline	8	≥ 16	2 - ≥ 16
	Piperacillin Tazobactam	4	≥ 128	1 - ≥ 128
	Tigecycline	1	2	0.5 - 4
	Amikacin	2	8	1 - 16
	Amoxicillin Clavulanic Acid	≥ 32	≥ 32	8 - ≥ 32
	Ampicillin	32	≥ 32	≤ 0.5 - ≥ 32
	Cefepime	≤ 0.5	≤ 0.5	≤ 0.5 - 1
	Ceftazidime	≤ 8	≤ 8	≤ 8
	Ceftriaxone	0.5	16	≤ 0.06 - 16
<i>S. marcescens</i>	Levofloxacin	1	4	0.25 - 8
	Meropenem	4	≥ 16	0.06 - ≥ 16
	Minocycline	8	8	4 - 16
	Piperacillin Tazobactam	1	8	0.25 - 32

Table 4. In vitro activity of tigecycline and comparative antimicrobial agents against other Gram negative clinical pathogens.

Microorganisms	Drugs	MIC 50	MIC 90	MIC Range
<i>A. baumannii</i>	Tigecycline	1	2	0.25 – 8
	Amikacin	≥64	≥64	≤0.5 - ≥64
	Amoxicillin Clavulanic Acid	≥32	≥32	≤8 - ≥32
	Ampicillin	≥32	≥32	8 - ≥32
	Cefepime	≥32	≥32	≤0.06 - ≥32
	Ceftazidime	≥32	≥32	≤8 - ≥32
	Ceftriaxone	≥64	≥64	4 - ≥64
	Levofloxacin	≥8	≥8	0.06 - ≥8
	Meropenem	≥16	≥16	0.12 - ≥16
	Minocycline	16	≥16	≤0.05 - ≥16
	Piperacillin Tazobactam	≥128	≥128	≤0.06 - ≥128
<i>P. aeruginosa</i>	Tigecycline	8	16	8 - ≥16
	Amikacin	8	≥64	2 - ≥64
	Amoxicillin Clavulanic Acid	≥32	≥32	≥32
	Ampicillin	≥32	≥32	≥32
	Cefepime	8	≥32	2 - ≥32
	Ceftazidime	16	≥32	≤8 - ≥32
	Ceftriaxone	≥64	≥64	64 - ≥64
	Levofloxacin	4	≥8	0.25 - ≥8
	Meropenem	4	16	≤0.06 - ≥16
	Minocycline	≥16	≥16	4 - ≥16
	Piperacillin Tazobactam	16	≥128	2 - ≥128
<i>H. influenzae</i>	Tigecycline	0.25	0.25	0.12 – 0.25
	Amikacin	2	4	2 – 8
	Amoxicillin Clavulanic Acid	1	2	0.25 - 2
	Ampicillin	≤0.5	4	≤0.5 - 8
	Cefepime	≤0.5	≤0.5	≤0.5
	Ceftazidime	≤8	≤8	≤8 - ≥32
	Ceftriaxone	≤0.06	0.5	≤0.06 - 16
	Levofloxacin	0.03	0.03	0.015 – 0.25
	Meropenem	≤0.06	0.25	≤0.06 – 0.25
	Minocycline	≤0.5	1	≤0.5 - 1
	Piperacillin Tazobactam	≤0.6	0.25	≤0.06 - 8

Table 5. In vitro activity of tigecycline and comparative antimicrobial agents against *Enterococcus spp.* and *Staphylococcus aureus*.

Microrganisms	Drugs	MIC 50	MIC 90	MIC Range
		µg/ L		
<i>Enterococcus spp.</i>	Tigecycline	0.12	0.25	0.03 – 0.5
	Amoxicillin/Clavulanic Acid	1	≥8	0.5 - ≥8
	Ampicillin	2	≥16	1 - ≥16
	Ceftriaxone	≥64	≥64	0.5 - ≥64
	Levofloxacin	≥32	≥32	0.5 - ≥32
	Linezolid	2	4	0.5 - 4
	Meropenem	8	≥16	4 - ≥16
	Minocycline	≥8	≥8	≤0.25 - ≥8
	Penicillin	8	≥8	2 - ≥8
	Piperacillin/Tazobactam	8	≥16	4 - ≥16
	Vancomycin	1	4	1 - ≥32
<i>S. aureus Met R</i>	Tigecycline	0.25	0.5	0.12 - 16
	Amoxicillin/Clavulanic Acid	8	≥8	0.12 - ≥8
	Ampicillin	≥16	≥16	≤0.06 - ≥16
	Ceftriaxone	64	≥64	4 - ≥64
	Levofloxacin	16	≥32	0.25 - ≥32
	Linezolid	4	4	≤0.5 - 4
	Meropenem	4	≥16	≤0.12 - ≥16
	Minocycline	0.5	≥8	≤0.25 - ≥8
	Penicillin	≥8	≥8	≤0.06 - ≥8
	Piperacillin/Tazobactam	8	≥16	1 - ≥16
	Vancomycin	1	2	1 - ≥32
<i>S. aureus Met S</i>	Tigecycline	0.12	0.5	0.12 - 2
	Amoxicillin/Clavulanic Acid	4	≥8	2 - ≥8
	Ampicillin	≥16	≥16	≥16
	Ceftriaxone	16	≥64	4 - ≥64
	Levofloxacin	16	32	0.25 - ≥32
	Linezolid	8	≥8	2 - ≥8
	Meropenem	1	≥16	≤0.12 - ≥16
	Minocycline	0.5	2	≤0.25 - 4
	Penicillin	≥8	≥8	≥8
	Piperacillin/Tazobactam	8	≥16	2 - ≥16
	Vancomycin	1	1	1

Table 6. In vitro activity of tigecycline and comparative antimicrobial agents against Streptococci.

Microrganisms	Drugs	MIC 50	MIC 90	MIC Range
		µg/ L		
<i>S. pneumoniae</i> PEN-R(5)	Tigecycline	1	1	1-2
	Amoxicillin/Clavulanic Acid	2	4	1 – 4
	Ampicillin	4	4	4
	Ceftriaxone	2	8	1-8
	Levofloxacin	1	1	1
	Linezolid	1	1	1
	Meropenem	1	1	0.5 - 1
	Minocycline	≥8	≥8	≥8
	Penicillin	4	4	2 - 4
	Piperacillin/Tazobactam	4	4	4 - 4
	Vancomycin	0.5	1	0.5 - 1
<i>S. pneumoniae</i> PEN-S(10)	Tigecycline	0.25	0.5	0.25 - 1
	Amoxicillin/Clavulanic Acid	≤0.03	≤0.03	≤0.03 – 0.12
	Ampicillin	≤0.06	≤0.06	≤0.06 – 0.25
	Ceftriaxone	≤0.03	≤0.03	≤0.03 - 1
	Levofloxacin	1	1	1
	Linezolid	≤0.5	1	≤0.5 - 1
	Meropenem	≤0.12	≤0.12	≤0.12
	Minocycline	2	8	0.25 - 8
	Penicillin	≤0.06	≤0.06	≤0.06 – 0.25
	Piperacillin/Tazobactam	≤0.25	≤0.25	≤0.25
	Vancomycin	0.5	0.5	0.25 – 0.5
<i>S. agalactiae</i>	Tigecycline	≤0.03	0.25	≤0.008 – 0.25
	Amoxicillin/Clavulanic Acid	0.06	0.06	≤0.03 – 0.06
	Ampicillin	≤0.06	≤0.06	≤0.06 – 0.12
	Ceftriaxone	0.06	0.06	≤0.03 - 1
	Levofloxacin	1	1	0.5 - 4
	Linezolid	≤0.5	1	≤0.5 - 2
	Meropenem	≤0.12	≤0.12	≤0.12
	Minocycline	2	≥8	≤0.25 - ≥8
	Penicillin	≤0.06	≤0.06	≤0.06
	Piperacillin/Tazobactam	≤0.25	≤0.25	≤0.25
	Vancomycin	2	0.5	≤0.12 – 0.5

REFERENCES

1. Anthony KB, et al. Clinical and microbiological outcomes of serious infections with multidrug-resistant Gram negative organisms treated with tigecycline. *Clin Infect Dis* 2008; 46: 567-70.
2. Bergeron J, Ammirati M, Danley D, et al. Glycylcyclines bind to the high-affinity tetracycline ribosomal binding site and evade Tet(M)- and Tet(O)-mediated ribosomal protection. *Antimicrob Agents Chemother* 1996; 40: 2226-8.
3. Bhattacharya M, Parakh A, Narang M. Tigecycline. *J Postgrad Med* 2009; 55: 65-8.
4. Breedt J, Teras J, Gardovskis J, et al. Tigecycline 305 cSSSI Study Group: Safety and efficacy of tigecycline in treatment of skin and skin structure infections: results of a double-blind phase 3 comparison study with vancomycin-aztreonam. *Antimicrob Agents Chemother* 2005; 49: 4658-66.
5. Bush K. Why it is important to continue antibacterial drug discovery. *ASM News* 2004; 70: 282-7.
6. Chopra I. Glycylcyclines: third-generation tetracycline antibiotics. *Curr Opin Pharmacol* 2001; 1: 464-9.
7. Fluit AC, Florijn A, Verhoef J, Milatovic D. Presence of tetracycline resistance determinants and susceptibility to tigecycline and minocycline. *Antimicrob Agents Chemother* 2005; 49: 1636-8.
8. Hanberger H, Garcia-Rodriguez JA, Gobernado M, Goossens H, Nilsson LE, Struelens MJ. Antibiotic susceptibility among aerobic Gram-negative bacilli in intensive care units in 5 European countries. French and Portuguese ICU Study Groups. *Jama* 1999; 281: 67-71.
9. Henwood CJ, Gatward T, Warner M, et al. Antibiotic resistance among clinical isolates of *Acinetobacter* in the UK, and *in vitro* evaluation of tigecycline (GAR-936). *J Antimicrob Chemother* 2002; 49: 479-87.
10. Livermore DM. Tigecycline: what is it, and where should it be used? *J Antimicrob Chemother* 2005; 56: 611-4.
11. Methods for dilution antimicrobial susceptibility tests for bacteria that grow aerobically; approved standard-eighth edition. CLSI document M07-A8. *Clinical Laboratory Standards Institute, Wayne, Pennsylvania*, 2010.
12. Oliva ME, Rekha A, Yellin A, et al. A multicenter trial of the efficacy and safety of tigecycline versus imipenem/cilastatin in patients with complicated intra-abdominal infections. *BMC Infect Dis* 2005; 5: 88.
13. Petersen PJ, Bradford PA, Weiss WJ, Murphy TM, Sum PE, Projan SJ. *In vitro* and *in vivo* activities of tigecycline (GAR-936), daptomycin, and comparative antimicrobial agents against glycopeptide-intermediate *Staphylococcus aureus* and other resistant Gram positive pathogens. *Antimicrob Agents Chemother* 2002; 46: 2595-601.
14. Reid GE, Grim SA, Aldeza CA, Janda WM, Clark NM. Rapid development of *Acinetobacter baumannii* resistance to tigecycline. *Pharmacotherapy* 2007; 27: 1198-201.
15. Rello J. Pharmacokinetics, pharmacodynamics, safety and tolerability of tigecycline. *J Chemother* 2005; 17 (S1): S12-S22.
16. Sacchidanand S, Penn RL, Embil JM, et al. Efficacy and safety of tigecycline monotherapy compared with vancomycin plus aztreonam in patients with complicated skin and skin structure infections: results from a phase 3, randomized, double-blind trial. *Int J Infect Dis* 2005; 9: 251-61.
17. Schafer JJ, Goff DA, Stevenson KB, Mangino JE. Early experience with tigecycline for ventilator-associated pneumonia and bacteremia caused by multidrug-resistant *Acinetobacter baumannii*. *Pharmacotherapy* 2007; 27: 980-7.
18. Shlaes DM, Projan SJ, Edwards JE. Antibiotic discovery: state of the state. *ASM News* 2004; 70: 275-81.
19. Wilcox MH. Efficacy of tigecycline in complicated skin and skin structure infections and complicated intra-abdominal infections. *J Chemother* 2005; 17 (S1): S23-S29.