

# Evaluation of the biochemical composition and antioxidant activity of preparation based on pigments extracted from the remaining biomass of *Arthrospira platensis*

Alina Beşliu,<sup>1,2</sup> Oleg Chiselîța,<sup>1</sup> Natalia Chiselîța,<sup>1</sup> Nadejda Efremova,<sup>1</sup> Tatiana Chiriac<sup>1</sup>

<sup>1</sup>Institute of Microbiology and Biotechnology of Technical University of Moldova, Fungal Biotechnology Laboratory, Chisinau; <sup>2</sup>Nicolae Testemitanu State University of Medicine and Pharmacy, Center of Advanced Biomedical Technologies (Core Research Facility), Chisinau, Republic of Moldova

## Abstract

Biotechnological research is currently focused on obtaining preparations based on natural pigments due to their properties and positive impact on human and animal health. Thus, this study

Correspondence: Alina Beşliu, Institute of Microbiology and Biotechnology of Technical University of Moldova, Fungal Biotechnology Laboratory, Chisinau, 1 Academiei Street, Republic of Moldova.  
E-mail: alina.besliu@imb.utm.md

Key words: *Arthrospira platensis*; antioxidant activity; carotenoid pigments; chlorophyll pigments; sulfated polysaccharides.

Acknowledgments: the results were obtained within Project 20.80009.5107.16, funded by the National Agency for Research and Development of Moldova.

Contributions: all authors contributed significantly to this work, including drafting and revising the manuscript.

Conflict of interest: the authors declare no conflict of interest.

Availability of data and materials: all data generated or analyzed during this study are included in this published article.

Ethics approval and consent to participate: not applicable.

Informed consent: not applicable.

Received: 21 April 2023.  
Accepted: 1 September 2023.  
Early view: 12 September 2023.

Publisher's note: all claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article or claim that may be made by its manufacturer is not guaranteed or endorsed by the publisher.

©Copyright: the Author(s), 2023  
Licensee PAGEPress, Italy  
Journal of Biological Research 2023; 96:11425  
doi:10.4081/jbr.2023.11425

This article is distributed under the terms of the Creative Commons Attribution-NonCommercial International License (CC BY-NC 4.0) which permits any noncommercial use, distribution, and reproduction in any medium, provided the original author(s) and source are credited.

aimed to evaluate the biochemical composition and antioxidant activity of the preparation based on pigments obtained from the remaining biomass of *Arthrospira platensis*. The obtained results established that the preparation is characterized by a high content of  $\beta$ -carotene, lutein, chlorophyll pigments, and sulfated polysaccharides. Due to its composition, the preparation also possesses high antioxidant activity and the catalase and superoxide dismutase enzymes. These findings highlight the high biological value of the new preparation and the enormous potential for implementation in medicine, the animal husbandry sector, and the food and cosmetic industry.

## Introduction

Currently, special attention to biotechnological research is directed toward the reuse of industrial secondary products obtained in enormous quantities following the manufacturing process.

The repurposing of microalgae *Arthrospira platensis* for deriving pigment-based formulations holds significant interest.

Owing to this, it possesses high biological activity and is commonly used as a nutritional supplement with a beneficial effect on human and animal health.<sup>1,2</sup> For the Republic of Moldova, the reuse of algal biomass remaining after the production of the BioR remedy is of importance. According to the specialized literature, the active part of BioR remedy preparation is composed of a combination of biologically active compounds (amino acids and oligopeptides, phospholipids, macro-, and microelements) that have been extracted from *Spirulina* biomass and purified. Clinical studies have demonstrated that the preparation exhibits antioxidant, cytoprotective, regenerating, immunomodulatory, anti-inflammatory, antiviral, hepatoprotective, and antiatherogenic activities.<sup>3,4</sup> However, after obtaining the preparation, the remaining biomass can be used for extraction of carotenoids, chlorophyll pigments, and sulfated polysaccharides which are of vital importance functioning primarily as aids in the photoprotection process. Carotenoid pigments have important metabolic functions, including conversion of vitamin A, enhancement of immune response, protection against eye diseases such as cataracts and macular degeneration, and cardiovascular diseases.<sup>5,6</sup> The major carotenoids of *Arthrospira platensis* are  $\beta$ -carotene and lutein which also have an important role in the antioxidant defense contributing to the protection of vulnerable biomolecules in the cell from the harmful effects of environmental stress, reactive oxygen species (ROS), and other aggressive chemical species.<sup>7-9</sup>

Another important pigment in the structure of the microalgae *Arthrospira platensis* is chlorophyll, a powerful antioxidant with

anti-inflammatory and anti-cancer properties. It is an essential additive and colorant in pharmaceutical, cosmetic, and food products.<sup>10,11</sup>

It was also established that the carotenoid and chlorophyll pigments obtained from *Arthrospira platensis* play an important role in zootechnics, being used to increase the productive and reproductive potential of animals. Their use contributes to the improvement of the quality of meat, eggs, and dairy products and offers several benefits for animals but also for human health.<sup>12</sup> According to some studies, carotenoid pigments contribute to the protection of the plasma membrane against peroxidation lipids and give membranes fluidity and flexibility that help sperm engage in the membrane fusion events associated with fertilization.<sup>13</sup>

Based on the above, the research aims to evaluate the biochemical composition and the antioxidant activity of the preparation based on pigments extracted from the remaining biomass of *Arthrospira platensis*.

## Materials and Methods

### Chemicals

The following reagents were used to determine the biochemical and antioxidant composition in the preparation based on pigments extracted from the remaining biomass of *Arthrospira platensis*: Total antioxidant activity was assessed utilizing 2,2'-azino-bis(3-ethylbenzothiazoline-6-sulfonic acid) (ABTS), generated by reacting equal volumes of 7 mM ABTS with 2.45 mM potassium persulfate (Sigma Aldrich Reagents, Darmstadt, Germany). This mixture was incubated in darkness for 12 hours to form the ABTS radical. The reaction mixture comprised 0.3 mL of pigment-based preparation and 2.7 mL of ABTS solution.

Catalase (CAT) activity was assessed using molybdenum as a catalyst and hydrogen peroxide 3% as the substrate for the reaction (Sigma Aldrich Reagents, Darmstadt, Germany).

The determination of superoxide dismutase (SOD) activity was carried out by assessing its capacity to inhibit the reduction of nitro-blue tetrazolium by superoxide in the presence of riboflavin (Sigma Aldrich Reagents, Darmstadt, Germany).

Protein extraction was carried out using 0.1N NaOH. Subsequently, 0.1 mL of the sample was collected, followed by the addition of 0.4 mL of distilled water and 2.0 mL of a mixture containing reagent A (sodium carbonate 2% in 0.1N sodium hydroxide) and reagent B (copper sulfate 0.5% in sodium citrate 1.0%). Finally, 0.2 mL of the Folin-Ciocalteu reagent was added and vigorously stirred (Sigma Aldrich Reagents, Darmstadt, Germany).

Total carbohydrates were determined using the anthrone reagent and D-glucose as a standard (Ecochimie, Chisinau, Republic of Moldova).

Sulfated polysaccharides were assessed by mixing 50 mL of alcian blue stock solution with 150 mL of 0.1N HCl and 1.5 mL of 96% ethanol containing 0.05M CaCl<sub>2</sub>. To the pectin precipitate, 2 mL alcian blue working solution and 7% CH<sub>3</sub>COOH were added.

The pigment extraction was performed using ethanol 96% (Eladam Pharma, Cojusna, Republic of Moldova).

### Process for obtaining the remaining biomass of *Arthrospira platensis*

The BioR remedy was obtained by cultivating *Arthrospira platensis* in a nutrient medium enriched with biochemical stimulators at a temperature of 30-35°C, under illumination of 18-24 thousand Erg/cm<sup>2</sup>/s and pH 8.5-10.0. The algal biomass was separated

through filtration, followed by the extraction, fractionation, and purification of the bioactive compounds.<sup>14</sup>

The biomass of *Arthrospira platensis* cyanobacteria, left over from the production of the BioR remedy, offered by the company «Ficotehfarm» LLC, Chisinau, Republic of Moldova, was used in the research.

### Process for obtaining the biologically active preparation based on pigments

Initially, the remaining *Arthrospira platensis* cyanobacteria biomass after BioR remedy production was dried at a temperature of 50±5°C until constant mass. Then, the biomass was subjected to grinding in an electric grinder for 3 minutes until it was transformed into a fine powder. In the next step, the biomass was mixed with 96% ethyl alcohol in a volume of 1:10. The obtained suspension was placed in a water bath with a thermostat at a temperature of 45°C for 30 minutes with periodic stirring. At the end of the process, the extract was separated from the biomass by centrifugation at 3500 rpm for 5 minutes. The preparation was concentrated by removing the ethyl alcohol in a rotary evaporator. Afterward, 0.1 mol/L phosphate buffer with pH 6.0 and 0.4 mmol/L EDTA were added to the preparation, and it was homogenized. The preparation was then standardized according to the dry substance up to a concentration of 40-44 mg/mL.

### Antioxidant activity

The total antioxidant activity was determined by the spectrophotometric method using the radical cation 2,2'-azino-bis 3-ethylbenzothiazoline-6-sulfonic acid (ABTS). The radical ABTS<sup>+</sup> is generated by the oxidation of ABTS with potassium persulfate and is reduced by the addition of hydrogen atoms.<sup>15</sup>

### Determination of pigment content

The β-carotene and lutein content was determined spectrophotometrically (Shimadzu UV-1280, Tokyo, Japan) using 96% ethyl alcohol, at room temperature, by shaking at 200 rpm for 30 minutes, separating the extract by centrifugation and determining the absorbance of the extract at the wavelength 450 nm.<sup>16,17</sup>

Supplementation of the preparation with chlorophyll pigments was carried out to improve the detoxifying and purifying properties of the preparation.

The content of chlorophyll pigments was determined by the spectrophotometric method, calculating the concentration of chlorophyll a, chlorophyll b, and the sum of chlorophylls for the ethanol solvent, according to the method described by Ritchie.<sup>18</sup>

### Determination of the biochemical composition

The protein content in the samples was determined according to the method described by Lowry *et al.*<sup>19</sup> The principle of the method is based on the formation of a copper complex with peptide bonds and its subsequent reduction in an alkaline medium.

The total carbohydrate content was determined spectrophotometrically at a wavelength of 620 nm using the anthrone reagent and D-glucose as standard.<sup>20</sup>

### Determination of sulfated polysaccharide content

To determine the content of sulfated polysaccharides, a method based on the binding of anionic carboxyl groups and sulfated ester groups of algal acid polysaccharides with alcian-blue dye was

used. This results in the formation of an insoluble precipitate. The optical density was measured at a wavelength of 610 nm. The difference between the absorbance of the dye in the blank sample and the absorbance of the dye remaining in the solution after precipitation was then calculated. This difference is proportional to the polysaccharide content in the sample, as described.<sup>21</sup>

## Evaluation of the antioxidant enzymes CAT and SOD

The activity of the CAT antioxidant enzyme was determined by the method that is based on the ability of hydrogen peroxide to interact with molybdenum salts, forming a stable-colored complex. The optical density was determined at a wavelength of 410 nm, as described.<sup>22</sup> SOD activity was determined by the method based on the inhibition of tetrazolium-nitro blue salt reduction in the presence of TEMED and riboflavin; the optical density was determined at a wavelength of 560 nm.<sup>23</sup>

## Statistical analysis

The statistical analysis of results was done using the statistical software kit Statistics 12. All experiments were performed in 3 replicates. Results were expressed by calculating the mean±standard deviation and confidence interval for a mean. All differences between values were considered statistically significant for  $p \leq 0.05$ .

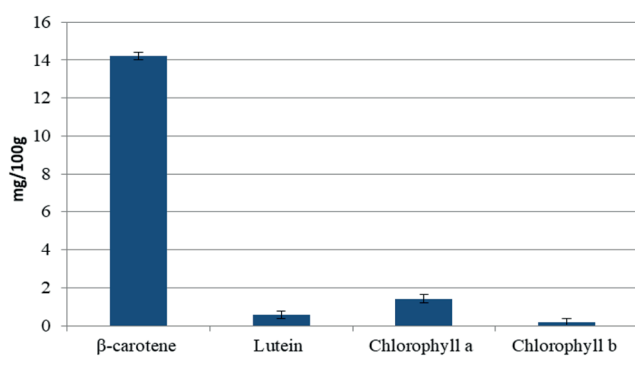
## Results

The preparation based on yellowish-green pigments, with a neutral smell, was obtained from the remaining biomass of *Arthrospira platensis* with a percentage yield of  $34.25 \pm 0.28\%$  d.w.

Initially, through spectrophotometric analysis in the preparation, the total content of carotenoid pigments was determined; the results obtained are shown in Figure 1.

The content of carotenoid pigments was  $14.77 \pm 3.93$  mg/100g, with a  $\beta$ -carotene content of  $14.21 \pm 0.02$  mg/100g. Additionally, the extract exhibited a well-balanced content of lutein which resulted in  $0.56 \pm 0.02$  mg/100g.

Subsequently, the concentration of chlorophyll pigments was evaluated, and the obtained results are illustrated in Figure 1. It was found that the preparation was characterized by a content of



**Figure 1.** Composition of carotenoid and chlorophyll pigments in the preparation obtained from the remaining biomass of *Arthrospira platensis*. Values are presented as mean±SD.

$1.42 \pm 0.07$  mg/100g of chlorophyll a and  $0.16 \pm 0.004$  mg/100g of chlorophyll b. The results of the biochemical composition of the preparation are presented in Table 1. According to the biochemical tests performed, it was determined that the preparation contains  $30.64 \pm 0.22\%$  d.w. proteins and  $28.44 \pm 0.05\%$  d.w. carbohydrates. Sulfated polysaccharides are an important component of the preparation. In this study yields of sulfated polysaccharides in the preparation showed a content of  $44.25 \pm 0.58$  mg/100g.

The result of the free radical capture efficiency of the preparation based on pigments obtained from the remaining biomass of *Arthrospira platensis* is presented in Table 1.

The reported value of  $195.94 \pm 9.15\%$  inhibition of the preparation demonstrates its strong ability to neutralize free radicals and oxidative species.

CAT and SOD are crucial antioxidant enzymes in algae, known for their role in defending against peroxidation activity and maintaining the redox state. In this study, the enzymatic activity of CAT and SOD was assessed, and the results are presented in Table 2. CAT activity in the experimental samples was  $1235 \pm 30.59$  mmol/min per mg of protein, while SOD activity was  $618 \pm 2.6$  U/mg protein.

The results demonstrate a high level of CAT enzyme's efficiency in breaking down harmful hydrogen peroxide into water and oxygen, preventing cellular damage from reactive oxygen species. The observed SOD activity indicates the enzyme's ability to convert superoxide radicals into less harmful forms, reducing oxidative damage. These findings highlight the potent antioxidant capacity of the pigment preparation obtained from the remaining biomass of *Arthrospira platensis*.

## Discussion

The spectrophotometric analysis revealed the presence of various pigments in the composition of the preparation obtained from the remaining biomass of *Arthrospira platensis*.

Our data on the composition of carotenoid pigments are in line with previous scientific research, which demonstrated that extracts obtained from *Arthrospira platensis* exhibit a higher content of  $\beta$ -carotene compared to other carotenoids. This finding indicates that the preparation is a safe and abundant source of  $\beta$ -carotene, making it suitable for consumption in large quantities for various production purposes.<sup>24</sup>

**Table 1.** Biochemical composition and antioxidant activity of the preparation based on pigments obtained from the remaining biomass of *Arthrospira platensis*.

Biochemical parameters	Value (mean±SD)
Protein content, % dry weight	$30.64 \pm 0.22$
Carbohydrates content, % dry weight	$28.44 \pm 0.05$
Sulfated polysaccharides content, mg/100g	$44.25 \pm 0.58$
Antioxidant activity, % inhibition	$195.94 \pm 9.15$

**Table 2.** Enzymatic activity of the preparation based on pigments obtained from the remaining biomass of *Arthrospira platensis*.

Enzymatic activity	Value (mean±SD)
CAT activity, mmol/min per mg	$1235 \pm 30.59$
SOD activity, U mg/protein	$618 \pm 2.6$

Another essential class of pigments found in the composition of the cyanobacterium *Arthrospira platensis* is chlorophyll pigments. These pigments serve as detoxifying and purifying phytonutrients that enhance the metabolism of carbohydrates, proteins, and lipids and have important antioxidant properties.<sup>25</sup>

The chlorophyll a content obtained in our research is in close agreement with the findings of Romero *et al.*, who reported a content of 11.08 µg/mL. However, there is a slight difference between the two studies regarding the content of chlorophyll b which could be attributed to variations in culture conditions and quantification methods. The growth environment, such as light intensity, temperature, nutrient availability, and pH, can influence chlorophyll synthesis in *Arthrospira platensis* cyanobacteria. Additionally, different quantification methods may have been employed, and variations in analytical techniques can lead to subtle differences in reported chlorophyll content.<sup>26</sup>

The results regarding the biochemical composition are in concordance with other studies in which it is described that the extracts obtained from *Arthrospira platensis* register a carbohydrate content that usually varies within the limits of 10 to 27% d.w.<sup>27,28</sup> Additionally, the studies presented by Abd El Baky and collaborators demonstrated that ethanol extraction from *Arthrospira platensis* gave the highest concentration of total carbohydrates.<sup>29</sup> The protein content cannot be compared with other studies because the previous extraction from the biomass was carried out for the production of the BioR remedy.

A significant advantage of the preparation is the presence of sulfated polysaccharides in its composition. Sulfated polysaccharides are heterogeneous complex natural polymers, featuring sugar units linked with sulfate groups with a different abundance depending on the species and extraction methods.<sup>29</sup>

They are of interest due to the complex group of macromolecules that give them a wide range of antioxidant, anticoagulant, antithrombotic, immunomodulatory, antiviral, and antibacterial biological properties.<sup>30-32</sup> They can also absorb toxic chemicals and play a major role as dietary fibers in maintaining animal and human health.<sup>33</sup> Similar results regarding obtaining sulfated polysaccharides from *Arthrospira platensis* have been reported in other studies, which gave high concentrations when extracted with alcohol due to the ability of alcohol to dissolve and separate the polysaccharides.<sup>29</sup>

The ABTS method was used in the research to determine the total antioxidant activity due to its ability to offer a comprehensive evaluation of the antioxidant power, along with its speed and stability.

Moreover, it is sensitive and can be used to measure both hydrophilic present in the preparation and lipophilic antioxidants such as carotenoid pigments.

The results of the evaluation of antioxidant activity showed the strong ability of the preparation based on pigments to neutralize free radicals and reactive oxygen species.

These results are due to the presence of pigments that significantly reduce oxidative stress that attacks and damages DNA, RNA, proteins, and lipids, leading to metabolic disturbances, tissue damage, and cell death.<sup>34,35</sup> It was established a positive correlation between the concentration of carotenoids in *Arthrospira platensis* biomass extracts and their radical scavenging activity, suggesting that these carotenoids are major contributors to the observed antioxidant properties.<sup>36,37</sup> The extracts obtained from *Arthrospira platensis* showed high antioxidant activity also in another study, in which a high content of pigments in the composition was recorded.<sup>38</sup> Furthermore, the antioxidative properties are determined by the presence of proteins, carbohydrates, and minerals within the composition.<sup>39</sup>

The high enzymatic activity is attributed to the unique bio-

chemical composition of the extract, wherein the combination of pigments and other biochemical components present in the structure plays a key role in enhancing enzymatic activity.

Finally, we should mention that the preparation based on pigments extracted from the biomass of *Arthrospira platensis* remaining from the production of the BioR remedy may offer significant advantages from an economic and environmental point of view. It helps reduce energy consumption and eliminates the need for cultivation and the application of many high-cost stimulation factors. The method aims to obtain the biomass extract with a more efficient approach, while also saving resources and has a lower cost compared to similar products.

## Conclusions

In this study, a new ecological and less expensive procedure was obtained to produce the preparation based on pigments from the remaining biomass of *Arthrospira platensis* cyanobacteria. The results demonstrate that the preparation is a good source of carotenoid pigments, chlorophylls, and sulfated polysaccharides. Due to its biochemical composition, the preparation also has high antioxidant and enzymatic activity.

## References

1. Wollina U, Voicu C, Gianfaldoni S, et al. *Arthrospira platensis* - potential in dermatology and beyond. *Open Access Maced J Med Sci* 2018;6:176-80.
2. Seyidoglu N, Galip N, Budak F, et al. The effects of *Spirulina platensis* (*Arthrospira platensis*) and *Saccharomyces cerevisiae* on the distribution and cytokine production of CD4+ and CD8+ T-lymphocytes in rabbits. *Austral J Vet Sci* 2017;49:185-90.
3. Rudic V. BioR. Biomedical and clinical studies. 1<sup>th</sup> ed. Chisinau: Elena V.I; 2007.
4. Rudic V, Cojocari A, Cepoi L, et al. Ficobiotehnologie-cercetare fundamentală și realizări practice [Phycobiotechnology-fundamental research and practical achievements] Chisinau: Elena-V.I 2007:365.
5. Krinsky NI, Johnson EJ. Carotenoid actions and their relation to health and disease. *Mol Aspects Med* 2005;26:459-516.
6. Mohan A, Misra N, Srivastav D, et al. *Spirulina* the nature's wonder: a review. *Lipids* 2019;5:7-12.
7. Solovchenko A, Chekanov K. Production of carotenoids using microalgae cultivated in photobioreactors. In: Paek K-Y, Murthy HN, Zhong J-J, eds. *Production of biomass and bioactive compounds using bioreactor technology*. 1<sup>th</sup> ed. Dordrecht: Springer; 2014. pp.63-91.
8. Mo NY. Extraction and determination of total carotenoids in orange heading Chinese cabbages. *J Northwest A&F Univ* 2014;42:206-14.
9. Wang L. Research on *Dunaliella* in  $\beta$ -carotene and sterol [dissertation]. Inner Mongolia: Inner Mongolia University 2014.
10. Ferruzzi MG, Blakeslee J. Digestion, absorption, and cancer preventative activity of dietary chlorophyll derivatives. *Nutr Res* 2007;27:1-12.
11. Hosikian A, Lim S, Halim R, Danquah MK. Chlorophyll extraction from microalgae: a review on the process engineering aspects. *Int J Chem Eng* 2010;11-8.
12. Saadaoui B, Mahmoudi M, Hassen HB, Rhouma M. Potential benefits of probiotics in livestock and poultry production. *J Anim Sci Technol* 2020;62:331-40.

13. Chethana S, Nayak CA, Madhusudhan MC, Raghavarao KSMC. Single step aqueous two-phase extraction for downstream processing of C-phycoerythrin from *Spirulina platensis*. *J Food Sci Technol* 2015;52:2415–21.
14. Iluta I, Rudic V, Chiriac T, et al. Preparatele LevoBioR — unguent, BioR-gel, BioR-loțiune, BioR-capsule: utilizarea lor în chirurgia maxilo-facială (buletin informativ) [LevoBioR preparations -ointment, gel BioR, lotion BioR, capsules BioR: their use in maxillofacial surgery (newsletter)]. *Medicina stomatologică* 2012;2:25-7.
15. Re R, Pellegrini N, Proteggente A, et al. Antioxidant activity applying an improved ABTS radical cation decolorization assay. *Free Radic Biol Med* 1999;26:1231-7.
16. Delia B, Rodriguez A. A guide to carotenoid analysis in food. SILSI Press. International Life Sciences Institute. One Thomas Circle, N.W. Washington, D.C. 2001:64.
17. Cepoi L. Photosynthetic pigments in *Porphyridium cruentum* under induced oxidative stress. *Akademios* 2014;4:116-20.
18. Ritchie RJ. Consistent sets of spectrophotometric chlorophyll equations for acetone, methanol and ethanol solvents. *Photosynth Res* 2008;96:166-17.
19. Lowry OH, Rosebrough N, Farr A, Randall R. Protein measurement with the Folin phenol reagent. *J Biol Chem* 1951;193:265-75.
20. Dey P, Harborne J. Methods in plant biochemistry. Carbohydrates. Academic Press 1993;2:529.
21. Zosim L, Bulimaga V, Rudic V, et al. Innovative process for increasing the content of acid polysaccharides in *Spirulina platensis* cyanobacterium. *Intellectus* 2019;1:139-42.
22. Komina A, Korostileva K, Gyrylova S, et al. Interaction between single nucleotide polymorphism in catalase gene and catalase activity under the conditions of oxidative stress. *Physiol Res* 2012;61:655-8.
23. Nekrasova GF, Kiseleva IS. Guide to laboratory and practical lessons. UMKD Ecological plant physiology. 2008, 157.
24. Mathew B; Sankaranaryanan R, Padmanabhan P. Evaluation of chemoprevention of oral cancer with *Spirulina fusiformis*. *Nutr Cancer* 1995;24:197-202.
25. Babadzhanov AS, Abdusamatoval N, Yusupova FM, et al. Chemical composition of *Spirulina platensis* cultivated in Uzbekistan. *Chem Nat Comp* 2004;40:340-4.
26. Romero L, Guevara M, Gómez B, et al. Production of pigments from *Arthrospira maxima* cultivated in photobioreactors. *Revista Colombiana de Biotecnología* 2017;19:108.
27. Christaki E, Bonos P, Florou P. Innovative microalgae pigments as functional ingredients in nutrition. *Handbook of marine microalgae: biotechnology advances*. Elsevier Inc 2015:223-43.
28. Gouveia L, Batista AP, Sousa I, et al. Microalgae in novel food products. *algae: nutrition*. *Environ Pollut Control* 2009:265-300.
29. Abd El Baky H, Hanaa El Baz KF, El-Latife SA. Induction of sulfated polysaccharides in *Spirulina platensis* as response to nitrogen concentration and its biological evaluation. *J Aquac Res Development* 2013;5:206.
30. Wang Z, Xie J, Shen M, et al. Sulfated modification of polysaccharides: synthesis, characterization and bioactivities. *Trends Food Sci Technol* 2018;74:147-57.
31. Nie X, Shi B, Ding Y, Tao W. Preparation of a chemically sulfated polysaccharide derived from *Grifola frondosa* and its potential biological activities. *Int J Biol Macromol* 2006;39:228-33.
32. Chaidedgumjorn A, Toyoda H, Woo ER, et al. Effect of (1→3)- and (1→4)-linkages of fully sulfated polysaccharides on their anticoagulant activity. *Carbohydr Res* 2002;337:925-33.
33. Manlusoc JKT, Hsieh CL, Hsieh CY, et al. Pharmacologic application potentials of sulfated polysaccharide from marine algae. *Polymers (Basel)* 2019;11:1163.
34. Lü JM, Lin PH, Yao Q, Chen C. Chemical and molecular mechanisms of antioxidants: experimental approaches and model systems. *J Cell Mol Med* 2010;14:840-60.
35. Upasani CD, Balaraman R. Protective effect of *Spirulina* on lead induced deleterious changes in the lipid peroxidation and endogenous antioxidants in rats. *Phytother Res* 2003;17:330-4.
36. Pérez-Gálvez A, Viera I, Roca M. Carotenoids and chlorophylls as antioxidants. *Antioxidants* 2020;9:505.
37. Jiménez-Escrig A, Jiménez-Jiménez I, Sánchez-Moreno C, Saura-Calixto F. Evaluation of free radical scavenging of dietary carotenoids by the stable radical 2,2-diphenyl-1-picrylhydrazyl. *J Sci Food Agric* 2000;80:1686-90.
38. Park WS, Kim HJ, Li M, et al. Two classes of pigments, carotenoids and C-phycoerythrin, in *Spirulina* powder and their antioxidant activities. *Molecules* 2018;23:2065.
39. Ezquerro-Brauer JM, Chan-Higuera JE. Capacidad antioxidante y mecanismo de acción de pigmentos en organismos marinos [Antioxidant capacity and mechanism of action of pigments in marine organisms]. *Ciencia UAT* 2021;15:186-97.