

Evaluation of the total antioxidant capacity of *Oliveria decumbens* and *Capparis spinosa*

Mohammad Darvishi,¹ Saade Abdalkareem Jasim,² Makhamadjalal I. Sarimsakov,³ Nadya J. Ibrahim,⁴ Suhad J. Hadi,⁴ Aymen Al-Sammarra'e,⁵ Roua Abulkassim,⁶ Noora M. Hameed,⁷ Waleed K. Alkhafaje,⁸ Zainab Hussein Adhab,⁹ Enas R. Alwaily,¹⁰ Mohadeseh Pirhadi,¹¹ Atieh Ganjeii,¹² Samira Shokri¹¹

¹School of Aerospace and Subaquatic Medicine, Beasat Hospital, AJA University of Medical Sciences, Tehran, Iran;

²Al-maarif University College, Medical Laboratory Techniques Department, Al-anbar-Ramadi, Iraq; ³Department of Pharmacology, Ferghana Medical Institute of Public Health, 2-a YangiTuron Street, Ferghana, Uzbekistan;

⁴Veterinary Medicine Collage, Al-Qasim Green University, Al-Qasim, Iraq; ⁵Department of Pharmacy, Al-Hadba University College, Iraq; ⁶Al-Manara College For Medical Sciences, Misan, Iraq; ⁷Anesthesia Techniques Department, Al-Nisour University College, Iraq; ⁸Anesthesia Techniques Department, Al-Mustaqbal University College, Babylon, Iraq; ⁹Department of Pharmacy, Al-Zahrawi University College, Karbala, Iraq; ¹⁰Microbiology Research Group, College of Pharmacy, Al-Ayen University, Thi-Qar, Iraq; ¹¹Department of Environmental Health Engineering, Division of Food Safety & Hygiene, School of Public Health, Tehran University of Medical Sciences, Tehran, Iran;

¹²Department of Food Science and Technology, Isfahan (Khorasgan) Branch, Islamic Azad University, Isfahan, Iran

Abstract

The anti-oxidative potential of extracts from two Iranian medicinal plants, *Oliveria decumbens* and *Capparis spinosa*, is

assessed in this study. The *Oliveria* genus is part of the Apiaceae family and contains only one species, *O. decumbens* Vent. Capers are either wild or cultivated plants that are found in many tropical and subtropical countries. *Oliveria decumbens* and *Capparis spinosa* aerial parts were dried and ground. The herbal solution was then created by combining herbal powder and methanol. Finally, the total antioxidant capacity of the plants was determined using the Ferric Iron Reducing Antioxidant Power (FRAP) assay and the ELISA reader at 570 nm. The total antioxidant capacities of *O. decumbens*, *C. spinosa* fruit, and *C. spinosa* leaves, respectively, were 3.82 mmol Fe²⁺/L, 1.96 mmol Fe²⁺/L, and 1.65 mmol Fe²⁺/L. These plants' essential oils are secondary metabolites that are widely used in the food, pharmaceutical, and health industries as antioxidant and antimicrobial compounds. In general, the traditional Iranian edible plants studied are good sources of natural antioxidant compounds.

Correspondence: Mohadeseh Pirhadi, Department of Environmental Health Engineering, Division of Food Safety & Hygiene, School of Public Health, Tehran University of Medical Sciences, Tehran, Iran. E-mail: m.pirhadi371@gmail.com

Key words: Antioxidant; medicinal plant; *Oliveria decumbens*; *Capparis spinosa* L.; treatment.

Conflict of interest: The authors declare no conflict of interest.

Availability of data and materials: All data generated or analyzed during this study are included in this published article.

Ethics approval and consent to participate: Not applicable.

Informed consent: Not applicable.

Received for publication: 30 May 2022.

Revision received: 19 September 2022.

Accepted for publication: 20 September 2022.

Publisher's note: All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article or claim that may be made by its manufacturer is not guaranteed or endorsed by the publisher.

©Copyright: the Author(s), 2022

Licensee PAGEPress, Italy

Journal of Biological Research 2022; 95:10644

doi:10.4081/jbr.2022.10644

This article is distributed under the terms of the Creative Commons Attribution-NonCommercial International License (CC BY-NC 4.0) which permits any noncommercial use, distribution, and reproduction in any medium, provided the original author(s) and source are credited.

Introduction

Oliveria decumbens Vent. is a fragrant annual herbaceous plant of the genus *Oliveria* in the Apiaceae family.¹ It can be found in Turkey, Iraq, and Syria, as well as in Iran's western and southwestern regions (Kermanshah, Chaharmahal Bakhtiari, Fars, Bushehr, and Khuzestan).² Botanical studies and medicinal uses of this plant in traditional Iranian medicine show that the medicinal plant *O. decumbens* has been used to treat indigestion, diarrhea, abdominal pain, and fever since ancient times in the western half of Iran.³

Capparis is a flowering and woody plant genus with a long root that penetrates deeply into the soil. Cabar, Kavar, Kavazeh, and Kavaz⁴ are the Iranian names for *Capparis spinosa*. *C. spinosa*, also known as Flinders rose, is a perennial plant with rounded, fleshy leaves and large white to pinkish-white flowers.⁵ The majority of the Capparaceae family is comprised of wild plants.^{6,7} Its fruits have traditionally been used to treat diabetes, headaches, fever, and rheumatism. *C. spinosa* roots, fruit, and bark have also been reported to be used as diuretics, tonics, and antimalarial

agents in Iranian traditional medicine.⁸ The content of 19-22% protein, 26% fiber, 1.7% ash, 31.6-36% oil source was reported.⁴ This plant's fruits and buds contain rutin and quercetin (phenolic and flavonoid compounds); rutin is a powerful antioxidant bioflavonoid.⁹

Tocopherol and sterol measurements in *C. spinosa* seed oil in Turkey revealed that it can be compared to other edible oils in terms of oxidative stability and antioxidant source.¹⁰ Many bioactive factors have been identified in this plant through phytochemical studies, including saccharides, glycosides, flavonoids, alkaloids, indoles, phenolic acids, terpenoids, essential oils, fatty acids, vitamins, and steroids.^{11,12} This plant's roots and flower buds contain pectin, saponin, aminoglycosides, and a substance known as caparirutin.⁵ Furthermore, in biological and chemical tests, aqueous-ethanolic extracts of the leaves demonstrated antioxidant activity.¹³

Traditional medicine relies heavily on medicinal plant extracts and active compounds for the majority of treatment.¹⁴ Anti-allergic, anti-inflammatory, antimicrobial, and antioxidant properties are among the many physiological benefits and properties of phenolic active compounds. These compounds are used in medicine, food, cosmetics, and agriculture;^{15,16} they also inhibit microbial activity and can kill germs without affecting consumer health.⁵ The use of plants in the treatment of diseases, particularly infectious diseases, has grown in recent years,¹ owing primarily to the negative side effects of synthetic medicinal compounds.¹⁷ Herbal derivatives, which are inspired by herbs, account for approximately 30 to 50 percent of all drugs available in pharmacies.¹⁸ The incorrect use of chemical drugs to treat infectious diseases has resulted in the emergence of resistant microbial isolates, the number of which is growing by the day.¹ Chemical-resistant strains necessitate efforts to discover new antimicrobial agents. Plants and their constituents, such as essential oils and plant extracts, have the potential to replace chemical drugs;¹⁹ however, the side effects of these compounds are less severe than those of chemical drugs.¹⁹ Previous research has shown that the essential oils of *O. decumbens* Vent. flowers and flowering branches, as well as the leaves and fruits of *C. spinosa* L., have potent antimicrobial properties.⁴ Because of the

growing trend among humans to use natural compounds derived from medicinal plant sources in the treatment and control of diseases, the antioxidant effect of medicinal plants *O. decumbens* Vent. and leaves and fruits of *C. spinosa* L. was investigated in this study.

Materials and Methods

Sampling and plant preparation

The aerial parts of *O. decumbens* and *C. spinosa* were collected from Dehloran city (Ilam Province, Southwest of Iran) in April 2022.

The plant was identified and approved in the Biotechnology and Medicinal Plants Research Center of Ilam University of Medical Sciences using the morphological keys from the book of plant flora of Ilam province.²⁰ Collected plants were cleaned and then dried in the open air in the darkness. The antioxidant testing solution was prepared using the method described by Dokhani *et al.*²¹

The characteristics of the medicinal plants used in this study are specified in Table 1.

Determination of the antioxidant activity of methanolic extract

The total antioxidant capacity of the plants was assessed by Ferric Iron Reducing Antioxidant Power (FRAP) assay of Naxifer kit.²¹

The amount of 2.2 mL of Reagent 2b (R2b) solution (Naxifer, Iran) was added to the parent R2a solution and vortexed until complete dissolution, obtaining the Reagent 2 (R2) solution. It was then mixed in a ratio of 1:1, again vortexed 5 times, and added to Reagent 1 (R1) solution. The resulting was the working solution of the antioxidant kit.²¹

Standard solutions from kit of 0, 0.2, 0.4, 0.6, 0.8 and 1 μ L were also prepared and added to microplate wells 1, 2, 3, 4, 5, and 6, respectively (well number 1 had no standard or zero concentration).²¹ The microplate was then incubated for 30 minutes at 35° C, and finally read at 570 nm with the Elisa reader.²¹

Table 1. Details of the utilized plants.

Scientific name	Plant family	Collection area	Geographical coordinates	Photo of the plant
<i>Oliveria decumbens</i> Vent.	Apiaceae	Dehloran	32° 41' 28" North, 47° 15' 58" East	
<i>Capparis spinosa</i> L.	Capparidaceae	Dehloran	32° 41' 28" North, 47° 15' 58" East	

Results

The results showed that the total antioxidant capacity was found as 3.82, 1.96 and 1.65 Fe²⁺/mmol for *Oliveria decumbens*, fruit of *Capparis spinosa* L. and leaves of *Capparis spinosa* L., respectively (Table 2).

Discussion

Chronic diseases are one of the most serious challenges confronting the world's health-care systems in the twenty-first century.^{22,23} They are diseases with a slow onset that are both progressive and long-term.^{24,25} Chronic diseases are caused by a variety of factors, including heredity, nutrition, environment, stress, and a variety of infectious and non-infectious factors.^{4,26} Non communicable diseases account for two-thirds of all deaths worldwide.²⁷

According to a World Health Organization report, approximately 80% of the world's population uses traditional medicine and methods to treat their diseases.²⁸ Medicinal plants are also used as guide compounds in drug synthesis.²⁸ In Asia, herbal medicines in the form of extracts are used by 40 to 62% of cancer patients. According to research on these patients, the benefits of using these herbal remedies outweigh the drawbacks.²⁹ Antioxidant activity is responsible for antioxidant activity in medicinal plants; antioxidants function by inactivating free radicals.³⁰

Our findings revealed that the total antioxidant capacity of *O. decumbens* was 3.82 mmol Fe²⁺/L. *O. decumbens* is a medicinal plant with potent antioxidant properties and a high free radical scavenging potential. According to Vazirzadeh *et al.*, administration of different *Oliveria decumbens* derivatives demonstrated remarkable antibacterial activity against streptococcosis, as well as improved antioxidant status and post-challenge immunity in Nile tilapia.³¹

The main compounds found in *O. decumbens* plants are thymol, carvacrol, paracetamol, and gamma-terpinene.³² Similar studies on the total antioxidant capacity of medicinal plants were conducted in various countries, for example, in Singapore in 2002, the antioxidant capacity of 28 types of fruit in *Bacopa caroliniana* in 2003, forty-five medicinal plants in Italy in 2003, 34 vegetables and 28 fruits.³³

C. spinosa L. flower is used in traditional medicine to treat liver and spleen diseases, as well as anemia and weakness.³⁴ Several studies³⁵ have confirmed its anti-diabetic, anti-cancer, and antioxidant properties. *C. spinosa* ameliorates cardiovascular disorders, liver damage, and nephropathy in diabetic animal models, according to Mazarei *et al.*³⁶ These results are attributed to its antioxidant phytochemicals such as phenolic compounds, flavonoids, carotenoids, tocopherol, and terpenes.

Table 2. Total antioxidant capacity of *O. decumbens* and *C. spinosa* L.

Common name	Scientific name	Total antioxidant capacity (mmol Fe ²⁺ /L)
Laeleh kohestan	<i>Oliveria decumbens</i>	3.82
Kabar fruit	<i>Capparis spinosa</i> L.	1.96
Kabar leaves	<i>Capparis spinosa</i> L.	1.65

Others, however, show that kabar leaves methanolic extract has antioxidant properties and a higher total phenol content than fruits and stems.^{9,37} The differences can be attributed to the plant's origin, method, operating conditions, and extraction solvent type. For normal metabolism, signal transmission, and cellular activity regulation, the human body requires oxidants, free radicals, and antioxidants.³⁷⁻⁴¹ Many diseases can be prevented by using medicinal plants and herbal antioxidants.⁴²⁻⁴⁷ According to Iranmanesh *et al.*, *C. spinosa* fruit extract is a potential source of natural antioxidants in the treatment of diseases.³⁴ The main compounds in Kabar root and fruit essential oils were methyl, isopropyl, and sec-butyl isothiocyanates, while the main compounds in leaf essential oil were thymol (24.6%) and isopropyl isothiocyanate (11%).³⁹ Because of its antioxidant properties, kabar fruit extract may protect against acetaminophen-induced hepatotoxicity.⁴⁸

Conclusions

The tested species appear to be valuable natural antioxidant sources with applications in both health care and the food industry. However, *in-vivo* safety and active compound identification must be thoroughly investigated before they can be used.

References

- Mahboubi M, Feizabadi M, Haghi G, et al. Antimicrobial activity and chemical composition of essential oil from *Oliveria decumbens* Vent. Iran J Med Aromatic Plants Res 2008;24:56-65.
- Amiri H, Lari Yazdi H, Dosti B, et al. Essential oil composition and anatomical study of *Oliveria decumbens* Vent. Iran J Med Aromatic Plants Res 2011;26:513-20.
- Bahraminejad S, Seifolahpour B, Amiri R. Antifungal effects of some medicinal and aromatic plant essential oils against *Alternaria solani*. J Crop Prot 2016;5:603-16.
- Saffarpour S, Givianrad M, Beheshti P. Detection and determination of antioxidant compounds of seed oil of *Capparis spinosa* L. in Iran. Iran J Med Aromatic Plants Res 2012;28:153-60.
- Safarzaei A, Sarhadi H, Haddad Khodaparast MH, et al. Optimization of aqueous and alcoholic extraction of phenolic and antioxidant compounds from caper (*Capparis spinosa* L.) Roots Assisted by Ultrasound Waves. Zahedan J Res Med Sci 2020;22.
- Gan L, Zhang C, Yin Y, et al. Anatomical adaptations of the xerophilous medicinal plant, *Capparis spinosa*, to drought conditions. Horticult Envir Biotechnol 2013;54:156-61.
- Giménez T, Mula D, Gea-Botella S, et al. Lipase catalyzed deacidification of tocopherol-rich distillates obtained from natural Vitamin E sources. Process Biochem 2019;77:70-6.
- Sozzi GO. Caper bush: botany and horticulture. Horticultural Rev 2001;27:125-88.
- Falih B, Mohammed S, Mohammed N. Effects of the silver nanoparticle synthesis from the leaves of the *Capparis spinosa* plant on the liver of mice infected with visceral leishmaniasis. Caspian J Environ Sci 2022;20:785-91.
- Mathhäus B, Özcan M. Glucosinolates and fatty acid, sterol, and tocopherol composition of seed oils from *Capparis spinosa* Var. *spinosa* and *Capparis ovata* Desf. Var. *canescens* (Coss.) Heywood. J Agric Food Chem 2005;53:7136-41.
- Moufid A, Farid O, Eddouks M. Pharmacological Properties

- of *Capparis spinosa* Linn. Int J Diabetol Vasc Dis Res 2015;3:99-104.
12. Zhou H, Jian R, Kang J, et al. Anti-inflammatory effects of caper (*Capparis spinosa* L.) fruit aqueous extract and the isolation of main phytochemicals. J Agric Food Chem 2010;58:12717-21.
 13. Mansour RB, Jilani IBH, Bouaziz M, et al. Phenolic contents and antioxidant activity of ethanolic extract of *Capparis spinosa*. Cytotechnol 2016;68:135-42.
 14. Obaid RF, Kadhimi Hindi NK, Kadhimi SA et al. Antibacterial activity, anti-adherence and anti-biofilm activities of plants extracts against *Aggregatibacter actinomycetemcomitans*: An in vitro study in Hilla City, Iraq. Caspian J Environ Sci 2022;20:367-72.
 15. Javed S, Javed I, Shoaib A, et al. Oleanolic acid (pentacyclic triterpenes) as a potential candidate for α -glycosidase inhibition activity. Adv Life Sci 2022;9:219-23.
 16. Aidi A, Bahmani M, Pirhadi M, et al. Phytochemical Analysis and Antimicrobial Effect of Essential Oil and Extract of *Loranthus europaeus* Jacq. on *Acinetobacter baumannii*, *Staphylococcus aureus*, and *Pseudomonas aeruginosa*. Kafkas Universitesi Veteriner Fakultesi Dergisi 2022;28.
 17. Zahrae Redouan F, Benitez G, Aboubakr B, et al. The status and perception of medicinal plants by local population of Talasemtane National Park (Northern Morocco). Caspian J Environ Sci 2020;18:131-47.
 18. Aslam M, Sial AA. Effect of hydroalcoholic extract of *Cydonia oblonga* miller (Quince) on sexual behaviour of wistar rats. Adv Pharmacol Sci 2014;2014.
 19. Srinivasan D, Nathan S, Suresh T, et al. Antimicrobial activity of certain Indian medicinal plants used in folkloric medicine. J Ethnopharmacol 2001;74:217-20.
 20. Mozaffarian VA. Flora of Ilam, Ilam, Iran. Contemporary Culture Publications: Tehran.2008; p. 936.
 21. Dokhani N, Nazer MR, Shokri S, et al. Determination and Evaluating the Antioxidant Properties of *Ziziphus nummularia* (Burm.f.) Wight & Arn., *Crataegus pontica* K.Koch and *Scrophularia striata* Boiss. Egyptian JVet Sci 2022;53:423-9.
 22. Jalalmanesh S, Darvishi M, Rahimi M, et al. Contamination of senior medical students' cell phones by nosocomial infections: A survey in a university-affiliated hospital in Tehran. Shiraz E Med J 2017;18:43920.
 23. Darvishi M, Nazer MR, Alipour MR. Investigating the end of patients suffering from diabetic foot hospitalized in Be'sat hospital of IRIAF from 2009 to 2014. Biomed Res 2017;28:4630-4633.
 24. Darvishi M, Forootan M, Nazer MR, et al. Nosocomial Infections, Challenges and Threats: A Review Article. Iranian JMed Microbiol 2020;14:162-81.
 25. Darvishi M. Antibiotic resistance pattern of uropathogenic methicillin-resistant *Staphylococcus aureus* isolated from immunosuppressive patients with pyelonephritis. J Pure Appl Microbiol 2016;10:2663-7.
 26. Tavakolpour S, Darvishi M, Mirsafaei HS, et al. Nucleoside/nucleotide analogues in the treatment of chronic hepatitis B infection during pregnancy: a systematic review. Infect Dis 2018;50:95-106.
 27. Darvishi M, Sadeghi SS. Evaluation of association of *Helicobacter pylori* infection and coronary heart disease (chd) among ccu patients. J Pure Appl Microbiol 2016;10:2621-6.
 28. Reyes-Jurado F, Cervantes-Rincón T, Bach H, et al. Antimicrobial activity of Mexican oregano (*Lippia berlandieri*), thyme (*Thymus vulgaris*), and mustard (*Brassica nigra*) essential oils in gaseous phase. Ind Crops Prod 2019;131:90-5.
 29. Zarabi S, Ahmadi S, Rostami R. Anticancer activity evaluation of methanolic extract of *Allium Jesdianum* and *Nectaroscordeum Coelzi* against HeLa and K562 cell lines. Yafte 2017;19.
 30. Rahmati M, Shokri S, Ahmadi M, et al. Comparison of pesticide effect of copper oxide nanoparticles synthesized by green chemistry and plant extracts on *Anopheles Stephensii* mosquitoes. Plant Biotechnol Persa 2022;4:89-96.
 31. Vazirzadeh A, Jalali S, Farhadi A. Antibacterial activity of *Oliveria decumbens* against *Streptococcus iniae* in Nile tilapia (*Oreochromis niloticus*) and its effects on serum and mucosal immunity and antioxidant status. Fish Shellfish Immunol 2019;94:407-16.
 32. Amin G, Sourmaghi MS, Zahedi M, et al. Essential oil composition and antimicrobial activity of *Oliveria decumbens*. Fitoterapia 2005;76:704-7.
 33. Zarban A, Malekaneh M, Hasanpour M, et al. Evaluation of antioxidant properties of 28 Iranian medicinal plants. Sci J Birjand Uni Med Sci 2004;11:9-15.
 34. Iranmanesh M, Najafi S, Yosefi M. Studies on Ethnobotany of important medicinal plants in Sistan. J Medicinal Herbs 2010;1:58-65.
 35. Tlili N, Elfalleh W, Saadaoui E, et al. The caper (*Capparis* L.): Ethnopharmacology, phytochemical and pharmacological properties. Fitoterapia 2011;82:93-101.
 36. Mazarei F, Jooyandeh H, Noshad M, et al. Polysaccharide of caper (*Capparis spinosa* L.) Leaf: Extraction optimization, antioxidant potential and antimicrobial activity. Int J Biolog Macromolecules 2017;95:224-31.
 37. Pham-Huy LA, He H, Pham-Huy C. Free radicals, antioxidants in disease and health. Int J Biomed Sci 2008;4:89-96.
 38. Afsharypour S, Jeiran K, Jazy AA. First investigation of the flavour profiles of the leaf, ripe fruit and root of *Capparis spinosa* var. *mucronifolia* from Iran. Pharmaceut Acta Helv 1998;72:307-9.
 39. Sanchooli M, Bagheri R, Jaber Sh, et al. Comparing the essential oil composition of root, leave and fruit of *Capparis spinosa* in field and habitat. Plant Ecosys 2012;8:27-40.
 40. Al-nuani RMA, Kadhimi NJ. The effect of *Capparis Spinosa* L. Plant on the cytochrome and glutathione to reduce the hepatotoxicity induced by paracetamol in mice. J Physics Conf Series 2020;1664:012121.
 41. Jafari H, Nemati M, Haddad Molayan P, et al. Scolicidal activity of *Mesobuthus eupeus* venom against the protoscolices of *Echinococcus granulosus*. Arch Razi Institute 2019;74:183-189.
 42. Masoudi R, Dadashpour Davachi N. Effect of Dietary Fish Oil on Semen Quality and Reproductive Performance of Iranian Zandi Rams. Arch Razi Institute 2021;76:621-9.
 43. Kamil Kadhimi Lawi Z, Ameen Merza F, Rabeeba Banoon S, et al. Mechanisms of Antioxidant Actions and their Role in many Human Diseases: A Review. J Chem Health Risks 2021;11:45-57.
 44. Durgawale PP, Patil MN, Joshi SA, et al. Studies on phytoconstituents, in vitro antioxidant, antibacterial, antiparasitic, antimicrobial, and anticancer potential of medicinal plant *Lasiosiphon eriocephalus* decne (Family: Thymelaeaceae). J Nat Sci Biol Med 2019;10:38.
 45. Halim S, Jasmi NA, Ridzuan PM, et al. Novel potential *Centella asiatica* extract in ameliorating neurotoxicity induced oxidative stress in chronic morphine dependant rat model. Int J Med Toxicol Legal Med 2020;23:79-83.

46. Othman Z, Khalep HRH, Abidin AZ, et al. The anti-angiogenic properties of *Morinda citrifolia*. L (Mengkudu) Leaves using chicken chorioallantoic membrane (CAM) assay. *Pharmacog J* 2019;11:1.
47. Solati K, Karimi M, Rafieian-Kopaei M, et al. Phytotherapy for wound healing: The most important herbal plants in wound healing based on Iranian ethnobotanical documents. *Mini-Rev Med Chem* 2020;21:500-19.
48. Alam M, Juraimi AS, Rafii M, et al. Evaluation of antioxidant compounds, antioxidant activities, and mineral composition of 13 collected purslane (*Portulaca oleracea* L.) accessions. *Bio Med Res Int* 2014;296063.

Non-commercial use only