

## Minimum bactericidal concentration of phenols extracted from oil vegetation water on spoilers, starters and food-borne bacteria

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### Abstract

The aim of the study was to assess the *in vitro* effect of phenols extracted from oil vegetation water (PEOW) on several food-borne strains. Antibacterial activity of PEOW was based on the minimum bactericidal concentration (MBC) on microtitre assay. The taxa tested were: *Staphylococcus* (n. 5), *Listeria* (n. 4), *Escherichia* (n. 2), *Salmonella* (n. 1), *Pseudomonas* (n. 3), *Lactobacillus* (n. 2) and *Pediococcus* (n. 1). *S. aureus* and *L. monocytogenes* showed the lowest level of resistance to PEOW (MBC=1.5-3 mg/mL). In contrast, the Gram negative strains (e.g. *S. Typhimurium* and *Pseudomonas* spp.) were in some cases unaffected by the tested doses and the MBCs ranged between 6 to 12 mg/mL. Starter cultures were dramatically reduced on growth (e.g. *Staphylococcus xylosum*; 0.75 mg/mL MBC). The thresholds for pathogenic strains could be considered for further applications of PEOW in food models (e.g. shelf life or challenge test studies).

### Introduction

Due to their bioactive properties, some vegetables or their extracts have been applied to a large number of human activities (e.g. food preservation) since ancient times (Medina *et al.*, 2006). In particular, a wide range of plant extracts showed certain levels of inhibition against bacterial growth. In most cases, the antibacterial activity is attributed to the presence of phenolic compounds that are metabolites involved in the resistance against parasites (Servili *et al.*, 2011a, 2011b).

Olives, virgin olive oil and its secondary products such as olive mill wastewater showed a high level of phenolic compounds. The gluco-

side oleuropein is the main phenolic compound (secoiridoids) in fruit; moreover in olive products the molecules of its hydrolysis exert a stronger antimicrobial activity (Medina *et al.*, 2006). However, most part of these compounds is lost in wastewater during the olive oil extraction process. It was recognized that the presence of phenolic compounds reduces the microbial degradability of oil vegetation waters (VWs) leading to pollution problems (Capasso *et al.*, 1995; Tafesh, *et al.*, 2011; Saadi *et al.*, 2007) However, VWs could be considered as additional resources for the virgin olive oil (VOO) industry. In fact several phenolic compounds occurring in VW, having the same chemical characteristics of VOO phenols, possess many biological properties that include antioxidant activity and, for this reason, may be used in food industry as natural preservatives or bio-active ingredients (Servili *et al.*, 2011b).

The aim of the study was to assess the *in vitro* bactericidal effect of purified phenols extracted from oil vegetation water (PEOW) obtained by membrane filtration techniques from VW on several food-borne strains (spoilage bacteria, food-borne pathogens and starter cultures). The assessment of bactericidal activity could define some threshold doses for a further application on real food models.

### Materials and Methods

In Figure 1 the protocol for the separation of the phenols extracted from oil vegetation water is reported.

A panel of 18 food-borne strains was investigated for the minimum bactericidal concentration (MBC; Table 1) in a microtiter assay. Several two-fold dilutions of the PEOW (Figure 1) were performed in a 20% ethanol/water solution (12.0; 6.0; 3.0; 1.5; 0.75; 0.375 mg/mL), considering a 65% of total phenolic content on the initial extract (Carraro *et al.*, 2014). Strains were grown overnight and then diluted to obtain a standardized inoculum. Each well was added with 50  $\mu$ L of PEOW and 200  $\mu$ L of bacterial suspension in a 96 well plate with a final bacterial concentration of 10<sup>5</sup> UFC/mL.

For each plate 3 wells without bacteria were performed according PEOW dilution (negative control). Moreover, for each strain 3 wells were added with 200  $\mu$ L of bacterial suspension and 50  $\mu$ L of a 20% ethanol/water solution; an additional positive control was performed for each strain using 250  $\mu$ L of bacterial suspension. Microtiter plates were incubated at specific temperature/time for each strains and the MBC was evaluated on agar plate by spreading 10  $\mu$ L of each suspension. The dose that kills the bacteria was recorded and the MBC was defined as the level that did not allowed the

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survival of all replicates. A visual examination of well bottoms was also applied in order to have a visible growth and define the minimum inhibitory concentration (MIC). Three different biological replicates were performed for each plate and all experiments were duplicates.

### Results

Table 1 reported the MBC value for the food-borne bacteria tested. Among pathogenic strains, *S. aureus* and *L. monocytogenes* showed the lowest level of resistance to PEOW (MBC=1.5-3 mg/mL). For Gram positive strains, some events of heteroresistance were observed and a stringent evaluation of MBC was based on the highest level that did not allowed the growth on all replicates. On the opposite, in Gram negative strains (e.g. *S. Typhimurium* and *Pseudomonas* spp.) the MBCs ranged between 6 to 12 mg/mL with the exception of *Escherichia coli* O:157 H7 (3 mg/mL). Starter cultures were dramatically reduced in growth (e.g. *S. xylosum*; MBC 0.75-1.5 mg/mL). *Pediococcus pentosaceus* seemed the more resistant species among acetic acid bacteria (LAB), the strain was able to survive at higher concentrations of PEOW (MBC 12 mg/mL). The ethanol/water solution did not affect the survival of any tested strains, the results were in agreement with the previously observation on *E. coli* (Carraro *et al.*, 2014). The final concentration of ethanol was around

1/4 of the initial solution (5%), this level was ineffective to kill the pre-inoculum of the tested bacteria.

The MIC is the lowest concentration of an antimicrobial that inhibit the growth of a microorganism. The visual examination has resulted in some misinterpretation due to the turbidity of the extract after the incubation, especially at higher PEOW concentrations (6-12 mg/mL); for this reason the evaluation of MIC is not reported.

## Discussion

As reported by Carraro *et al.* (2014) the composition of the extract showed that Oleuropein-aglycone di-aldehyde was the major secoiridoid constituent ( $471.7 \pm 1.9$  mg/g). Others chemical compounds are Hydroxytyrosol ( $72.7 \pm 0.6$  mg/g), Tyrosol ( $17.8 \pm 0.1$  mg/g) and Verbascoside ( $83.6 \pm 1.0$  mg/g). The bactericidal effect was linked to a combination of these substances. The *in vitro* tests suggested an interesting bactericidal effect of PEOW, especially on Gram positive food-borne pathogens. Other studies reported that *S. aureus* and *L. monocytogenes* showed a higher sensitivity to phenols derived from several olive matrices (*e.g.* olive oil, olive leaf and purified compounds) (Medina *et al.*, 2006; Pereira *et al.*, 2007). Moreover, Thafesh *et al.* (2011) suggested that the use of a combina-

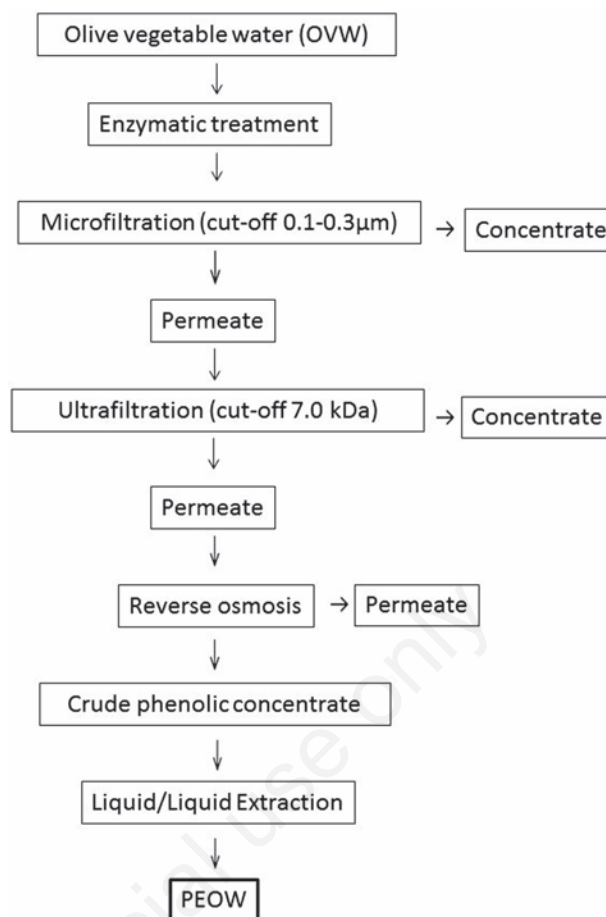


Figure 1. Protocol for the separation of the phenols extracted from oil vegetation water from olive vegetable water. Based on Servili *et al.* (2011a).

Table 1. Species, origin and growth mediums of the strains tested in the study and their relative minimum bactericidal concentration.

Species	Origin	Medium	Temperature (°C) - incubation (h)	MBC (mg/mL)
<i>Staphylococcus aureus</i> ATCC 29213	Wound	LB and agar	37 - 24	1.5
<i>Staphylococcus aureus</i> LMG 8224	Clinic isolate	LB and agar	37 - 24	3
<i>Staphylococcus aureus</i> 415*	Salami	LB and agar	37 - 24	1.5
<i>Listeria monocytogenes</i> 21962 AL 8/4	Food	LB and agar	37 - 24	3
<i>Listeria monocytogenes</i> 1271*	Salami	LB and agar	37 - 24	3
<i>Listeria monocytogenes</i> LMG 13305	Fresh cheese	LB and agar	37 - 24	3
<i>Listeria innocua</i> ATCC 33090	Bovine brain	LB and agar	37 - 24	1.5
<i>Escherichia coli</i> O:157 H7 NCTC 12900	Human diarrhea	LB and agar	37 - 24	3
<i>Escherichia coli</i> LMG 8223	Clinic isolate	LB and agar	37 - 24	6
<i>Salmonella</i> Typhimurium ATCC 14028	Liver of hen	LB and agar	37 - 24	12
<i>Pseudomonas fluorescens</i> MOZ3°	Mozzarella cheese	LB and agar	22 - 48	6
<i>Pseudomonas fluorescens</i> PS22#	Mozzarella cheese	LB and agar	22 - 48	6
<i>Pseudomonas aeruginosa</i> 4C2T°	Stracchino cheese	LB and agar	37 - 24	6
<i>Staphylococcus xylosum</i> 100M SA1- (3)§	Starter for salami	LB and agar	30 - 48	0.75
<i>Staphylococcus xylosum</i> SA1-100M (7)§	Starter for salami	LB and agar	30 - 48	1.5
<i>Lactobacillus curvatus</i> SA1-100M (13)§	Starter for salami	MRSB/agar	30 - 48	1.5
<i>Lactobacillus curvatus</i> SA1-100M (14)§	Starter for salami	MRSB/agar	30 - 48	1.5
<i>Pediococcus pentosaceus</i> 923 CECT	Fermented milk	MRSB/agar	30 - 48	12

MBC, minimum bactericidal concentration; LB, lysogeny broth. \*Isolated by VenetoAgricoltura, Thiene (VI); °isolated by University of Udine; #isolated by Institute for Experimental Veterinary Medicine of Venice; §isolated by Bioagro srl, Thiene (VI).

tion of polyphenols extracted by olive mill wastewater is effective against several human pathogens. The present results confirmed the potential of PEOW, though the bioactivity could be related to the content of certain specific phenolic compounds (Thafesh *et al.*, 2011). As noted, the visual examination of MIC failed at higher concentrations of PEOW. This is probably due to the formation of a brown-coloured quinonic form due to the oxidative degradation of phenols. The present results suggested two thresholds of bactericidal effect on Gram positive food-borne pathogens. However, other pertinent considerations need to be considered when these products are added to the food (*e.g.* organoleptic traits, diffusion, bonding with some food constituents).

## Conclusions

Taking into account MBC results, the LAB and *S. xylosum* are among the most sensitive bacteria. In a functional milk beverage fortified with phenolic compounds Servili *et al.* (2011b) did not find a significant reduction in LAB growth. Still, the level of inclusion was limited (100 and 200 mg/L). In other fermented products, the supplementation of higher levels of PEOW, as a natural ingredient, could reduce the performance or the quality of ripening. For further applications, a deep screening of starter cultures is required in order to select

the species able to grow in presence of phenol during fermentation. The MBC suggested that one strain of *Pediococcus pentosaceus* was able to grow at 12 mg/mL: this species could be applied to some fermented food (*e.g.* salami). These results highlighted the potential of PEOW against some food-borne bacteria. This work is the first step before the use of these substances on the food models (*e.g.* challenge test and storage test) for the further application of this extract as an ingredient. The two thresholds proposed for the pathogenic food-borne bacteria need to be considered together with other pertinent food aspects (*e.g.* organoleptic traits, antioxidant effects).

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