## REVIEW

# The influence of oral antioxidants on men with infertility: A systemic review

Tamer A. Abouelgreed <sup>1</sup>, Mohamed A. Amer <sup>2</sup>, Hassan Mamdouh <sup>2</sup>, Ahmed F. El-Sherbiny <sup>3</sup>, Hany Aboelwafa <sup>2</sup>, Sameh F. Fahmy <sup>2</sup>, Omar A. Omar <sup>2</sup>, Mohammed Abdelshakour <sup>2</sup>, Mohammad Elesawy <sup>2</sup>, Mohamed Sonbol <sup>2</sup>, Ahmed N. Maawad <sup>2</sup>, Osama K. Elsayed <sup>2</sup>

<sup>1</sup> Department of Urology, Al-Azhar University, Cairo, Egypt;

<sup>2</sup> Department of Dermatology & Andrology, Al-Azhar University, Cairo, Egypt;

<sup>3</sup> Department of Andrology, International Islamic Center for Population Studies and Research, Al-Azhar University, Cairo, Egypt.

**Summary** Objective: This study aims to investigate the current evidence regarding the impact of oral antioxidant supplementation on semen parameters of infertile men.

Materials and methods: We conducted a systematic search of PubMed, and Cochrane electronic databases, adhering to modified Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA) guidelines. The focus was on studies exploring the effects of antioxidant therapy on infertile men, with an examination of antioxidants in terms of types, doses, rationale for use, and their impact on semen parameters measures.

Results: A total of 18 studies that met the inclusion criteria were included in this study. Out of these, 14 studies reported a significantly positive influence of antioxidant therapy on basic semen parameters and advanced sperm function. These comprised 11 randomized clinical trials and 7 prospective studies. Commonly utilized antioxidants included Vitamin E, Vitamin C, carnitines, co-enzyme Q10, N-acetyl cysteine, zinc, selenium, folic acid, and lycopene.

Conclusions: Overall, antioxidants generally demonstrate a favorable effect on semen parameters of infertile men. However, further research is necessary to pinpoint the optimal antioxidant regimen that can be applied safely and effectively in clinical practice.

KEY WORDS: Infertility; Antioxidants; Semen parameters.

Submitted 30 January 2024; Accepted 18 February 2024

#### INTRODUCTION

Infertility is defined as the inability to conceive after at least 12 months of regular, unprotected intercourse, affecting approximately 15% of couples globally. Notably, male factors contribute to nearly half of the reported cases, often linked to disturbances in testicular function or blockage of reproductive passages (1, 2). Consequently to the efforts to unravel the molecular-level of idiopathic male infertility, the substantial role played by *oxidative stress* (OS) has been underscored. OS refers to the imbalance in the body's redox state, arising from either excessive oxidants or insufficient antioxidants. *Reactive oxygen species* (ROS), commonly known as 'free radicals,' are highly reactive oxygen-derived molecules characterized by unpaired electrons in their outer valence orbital. These include oxygen-centered radicals (hydroxyl radical, nitric oxide radical, and superoxide anion radical) and non-radical derivatives (hydrogen peroxide, peroxynitrite anion, and hypochlorous acid) (3, 4). Various endogenous (e.g., immature spermatozoa, leukocytes, varicocele) and exogenous (e.g., testicular hyperthermia, environmental and habitual exposures) factors have been identified as potential causes of increased ROS production. To counterbalance ROS, antioxidants play a crucial role in maintaining the desired redox equilibrium for optimal sperm function (5). Seminal fluid is rich in antioxidants that nourish and protect sperm, existing in two forms: enzymatic and non-enzymatic antioxidant systems (6). The enzymatic system comprises naturally occurring antioxidants, including glutathione peroxidase, superoxide dismutase, and catalase, believed to originate from the prostate and found in sperm cells or seminal plasma. In contrast, the non-enzymatic system consists of various compounds obtained through diet or supplements. When an excess of ROS is produced or antioxidant activity is insufficient, OS occurs, disrupting the equilibrium between oxidation and reduction. Spermatozoa are particularly susceptible to OS due to their low levels of enzymatic antioxidants, originating from the prostate. Additionally, the high concentration of polyunsaturated fatty acids, notably docosahexaenoic acid, in the sperm cell's plasma membrane makes them attractive targets for ROS-induced oxidation reactions. Recent decades have witnessed significant progress in understanding male infertility, incorporating tests like sperm DNA fragmentation (SDF) and measures of OS to enhance clinicians' insights into male fertility potential (7,8). Advances in assisted reproductive therapy (ART) have allowed previously infertile men to father biological children. However, OS remains a critical factor influencing reproductive outcomes, both in natural conception and with ART. Approximately 25% of infertile men exhibit significant levels of ROS in their semen compared to fertile counterparts (9). OS negatively impacts semen parameters, fertilization rates, embryonic development, and pregnancy rates (10, 11).

The impact of antioxidants on fertility depends by substantial variations in antioxidant forms, dosages, combinations, and outcome measures across studies. This literature review aims to explore the most commonly used antioxidants in treating male infertility and investigate the effect of their doses that may confer benefits on basic semen parameters, advanced sperm function tests, outcomes of ART, and live-birth rates.

#### **MATERIALS AND METHODS**

#### Research strategy

The research strategy adhered to modified *Preferred Reporting Items for Systematic Reviews and Meta-Analyses* (PRISMA) guidelines (12). A literature search was conducted on PubMed, and Cochrane electronic databases to identify studies exploring the effectiveness of antioxidants in treating male infertility. Keywords and Medical Subject Heading (MeSH) phrases included 'antioxidants,' 'male infertility,' semen parameters'.

#### Study selection

Screening of articles was performed based on title and abstract by all authors followed by examination of relevant full papers. Additionally, review articles were scrutinized for potential inclusion (male patients, human studies). Exclusion criteria were based on study methods (retrospective, case report, editorial, or commentary). Data extraction was cross-checked and verified.

#### Outcome measures

The outcomes of interest included the type and dosage of antioxidants, their mechanism of action, impact on basic semen parameters and advanced sperm function tests.

## RESULTS

The search strategy identified 380 articles, with 315 excluded based on title and/or abstract. The remaining 65 articles underwent screening, leading to the identification of 18 studies that met the inclusion and exclusion criteria. These comprised 11 randomized clinical trials (13-23) and 7 prospective studies (24-30). Out of these, 14

#### Table 1.

The mode of action of commonly used antioxidants.

Antioxidant compound	Mechanism of action	
Ascorbic acid (vitamin C)	Neutralises free radicals	
Tocopherol (vitamin E)	Neutralises free radicals	
Folate (vitamin B9) Selenium	Scavenges free radicals	
Selenium	Enhancement of enzymatic antioxidant activity	
Zinc	Inhibition of NADPH oxidase	
Carnitines	Neutralizes free radicals and acts as an energy source	
CoQ10	In its reduced form, scavenges free radicals intermediate in mitochondrial electron transport system	
NAC	Enhances enzymatic antioxidant activity	
Lycopene	Quenches free radicals	

#### Table 2.

Studied antioxidants effect on semen parameters.

Clinical circumstance	Antioxidant	Reference
Basic semen parameters		
Oligozoospermia	Vitamin E, vitamin C, NAC, carnitines, CoQ10, lycopene, selenium and zinc	
	Vitamin E (300 mg)	(12)
	Vitamin E (180 mg), vitamin A (30 mg) and essential fatty acids or NAC (600 mg)	(25)
	NAC (600 mg) + other vitamins/minerals	(13)
	LC (2 g)	(14)
	CoQ10 (300 mg)	(15)
	NAC (600 mg) and selenium (200 mg)	(16)
	Folic acid (5 mg) + zinc (66 mg)	(17)
	Lycopene (2 mg)	(26)
Asthenozoospermia	Vitamin E, vitamin C, NAC, carnitines, CoQ10, lycopene, selenium and zinc	
	Vitamin E (400 mg) + selenium (200 lg)	(27)
	CoQ10 (300 mg)	(15)
	Lycopene (2 mg)	(26)
	LC (2 g) and LAC (1 g)	(19)
	NAC (600 mg)	(20)
	NAC (600 mg) and selenium (200 mg)	(16)
	Zinc (400 mg), vitamin E (20 mg) and vitamin C (10 mg)	(18)
Teratozoospermia	Vitamin E, NAC, lycopene, selenium and zinc	
	Vitamin E (400 mg) + selenium (200 lg)	(27)
	Lycopene (8 mg)	(28)
	NAC (600 mg) and selenium (200 mg)	(16)
	Zinc (400 mg), vitamin E (20 mg) and vitamin C (10 mg)	(18)
Advanced sperm function		
OS (oxidative stress)	Vitamin E, vitamin C, NAC, selenium and zinc	
	Vitamin E (300 mg)	(22)
	Vitamin E (180 mg) and b-carotene (30 mg)	(25)
	Vitamin E (20 mg), vitamin C (10 mg) and zinc (400 mg)	(18)
	Vitamin E (400 mg) and selenium (225 g)	(23)
	NAC (600 mg)	(20)
High SDF	Vitamin E, vitamin C, zinc, selenium and folic acid	
(sperm DNA fragmentation)	Vitamin E (1 g) + vitamin C (1 g) Vitamin C (400 mg), vitamin E (400 mg), b-carotene	(21)
	(18 mg), zinc (500 mmol) and selenium (1 mmol) LC (1500 mg); vitamin C (60 mg); CoQ10 (20 mg);	(29)
	vitamin E (10 mg); zinc (10 mg); folic acid (200 lg)	(30)
	( 0), (	()

studies (13-19, 24-30) reported a significant positive effect of antioxidant therapy on various parameters such as basic semen parameters, advanced sperm function tests. Commonly investigated antioxidants included vitamin E, vitamin C, carnitines, *N-acetyl cysteine* (NAC), *co-enzyme Q10* (CoQ10), zinc, selenium, folic acid, and lycopene. The doses and mechanisms of action for each antioxidant are presented in Table 1. Additionally, Table 2 outlines the outcomes of antioxidant treatment that were assessed across studies (13-30).

## DISCUSSION

Antioxidants, whether biological or chemical compounds, act scavenging free radicals, neutralizing their effects, and disrupting the chain reaction leading to OS in body tissues. In the context of male fertility, antioxidants are commonly prescribed for their accessibility and relatively low cost. However, conflicting results have been observed in studies assessing the impact of antioxidant therapy on male fertility. While some studies reported positive effects on semen parameters, sperm function, and pregnancy rates, others failed to confirm such benefits or even indicated a negative influence on male fertility. The heterogeneity across studies and the unknown optimal balance of the redox system for sperm function contribute to these discrepancies. Overconsumption of antioxidants may lead to reductive stress with potential detrimental effects on human health, including impairment of mitochondrial activity (31-33).

## Antioxidants: Mechanism of action and rationale for use

Numerous compounds with antioxidant properties have been explored for treating male infertility (Table 1). Understanding the mechanisms of action of commonly used compounds is crucial before delving into the associated evidence in clinical practice.

Vitamin E ( $\alpha$ -tocopherol): This potent chain-breaking antioxidant is a fat-soluble compound predominantly located in cell membranes. It quenches free hydroxyl radicals and superoxide anions, reducing lipid peroxidation initiated by ROS at the plasma membrane level. Vitamin E levels found to be correlated to the percentage of motile spermatozoa in semen. Lower vitamin E levels were observed in the semen of infertile men (34).

*Vitamin C (ascorbic acid)*: A water-soluble compound that is found in high concentrations in seminal plasma more than in blood serum. It neutralizes hydroxyl, superoxide, and hydrogen peroxide radicals, offering protection against endogenous oxidative damage. Seminal fluid analyses from infertile men with asthenozoospermia revealed lower vitamin C levels and higher ROS levels compared to fertile controls (35).

*Carnitines (L-carnitine and L-acetyl carnitine)*: These watersoluble antioxidants are involved in sperm metabolism, fueling essential activities like sperm motility. Carnitines exhibit antioxidant activities by scavenging superoxide anions and hydrogen peroxide radicals, inhibiting lipid peroxidation. Semen samples from infertile men with oligoasthenoteratozoospermia showed significantly lower carnitine levels (36).

*CoQ10*: This vital antioxidant is present in almost all body tissues, particularly in sperm mitochondria involved in cellular respiration and energy production. CoQ10's role in promoting motility and acting as an antioxidant is rationalized by its inhibitory effect on superoxide formation (37).

*NAC (N-acetyl cysteine)*: This amino acid, converted to cysteine in body tissues, acts as a precursor of glutathione, a crucial naturally occurring antioxidant that neutralizes various ROS. NAC directly reduces OS by scavenging hypochlorous acid and hydroxyl radicals. Studies have documented its positive influence on germ cell survival, showcasing reductions in ROS levels and improvements in sperm motility after incubation with *NAC (38)*.

Selenium: An essential trace element, selenium's role in spermatogenesis is linked to its ability to protect sperm

DNA against OS damage. Selenium's antioxidant properties are associated with its augmentation of glutathione function. Selenoenzymes, including *phospholipid hydroperoxide glutathione peroxidase* (PHGPX) and sperm capsular selenoprotein glutathione peroxidase, contribute to maintaining sperm structural integrity. Selenium deficiency is often correlated with morphological sperm abnormalities and impaired motility (39).

Zinc: Another essential trace element, zinc, plays vital roles in RNA and DNA metabolism, signal transduction, gene expression, and apoptosis regulation. Its antioxidant properties stem from its ability to decrease the production of hydrogen peroxide and hydroxyl radicals by antagonizing redox-active transition metals like iron and copper. Higher zinc concentrations in seminal plasma are observed in fertile men compared to subfertile men, and zinc deficiency is associated with various sperm structural abnormalities (40).

*Folic Acid (Vitamin B9)*: Involved in nucleic acid synthesis and amino acid metabolism, folic acid is used in male infertility treatment for its free radical scavenging abilities. Folic acid intake is linked to an increased reduced-to-oxidized glutathione ratio (41).

*Lycopene*: A naturally synthesized carotenoid found in fruits and vegetables, lycopene contributes significantly to the human redox defense system due to its potent ROS quenching abilities (42).

## Antioxidant effect on basic semen parameters

Semen analysis remains a fundamental test for assessing male fertility due to its simplicity and wide availability. However, continuous updates in reference values pose challenges in interpreting the evidence surrounding the potential impact of antioxidants, as changes in criteria may label patients differently. Despite these challenges, studies have reported improvements in basic semen parameters following oral antioxidant intake, either alone or in combination.

*Vitamin E*: Used in combination with other vitamins and minerals, vitamin E alone (300 mg daily) showed a significant improvement in sperm motility in infertile men (23). A study comparing vitamin E, clomiphene citrate, and a combination of both treatments in patients with idiopathic oligoasthenozoospermia reported a significant improvement in sperm concentration and motility with the combined regimen (13). Another study using vitamin E (400 mg) + selenium (200 µg) for 100 days showed a significant improvement in sperm motility, morphology, or both in infertile men (34). However, some studies failed to reproduce significant effects on semen parameters using vitamin E alone or in combination with other antioxidants (28, 30).

*Vitamin C*: Studies demonstrated the positive effects of vitamin C, particularly in heavy smokers, showing dose-dependent improvements in sperm quality (43). Vitamin C as an adjunct therapy post-varicocelectomy resulted in a statistically significant improvement in sperm motility and morphology compared to a placebo group (44). Several antioxidant supplements containing vitamin C have been investigated, showing significant improvement in sperm motility with combinations including zinc and vitamin E (19).

*Carnitines*: Studies confirmed the significant influence of carnitines, especially on sperm motility (45). A placebocontrolled trial demonstrated significant improvement in all semen parameters, with the most significant increase in sperm motility, using a combined treatment of L-carnitine (2 g) and L-acetyl carnitine (1 g) (20). LC and LAC treatment showed significant improvement in semen parameters, particularly in patients with lower baseline values of motility (46).

*CoQ10*: CoQ10 significantly improved sperm concentration and motility compared to placebo in men with idiopathic oligoasthenozoospermia (16). A clinical trial demonstrated improvements in sperm morphology, catalase, and superoxide dismutase with CoQ10 treatment (47). These findings highlight the potential benefits of antioxidant supplementation in improving sperm parameters, but variations in study outcomes emphasize the need for further research and standardization. A systematic review of three randomized controlled clinical trials involving 332 infertile men indicated that CoQ10 treatment (200-300 mg daily) led to a significant increase in sperm concentration (MD 5.33 x 10^6 sperm/mL, p < 0.001) and motility (MD 4.5%, p < 0.001) (48).

NAC (*N*-acetyl cysteine): In a randomized placebo-controlled study of 120 patients with idiopathic infertility, daily treatment with 600 mg NAC for 3 months resulted in a significant improvement in volume, motility, and viscosity of semen compared to placebo (21). Combining 600 mg NAC with 200 mg selenium showed a significant improvement in all semen parameters, with a dosedependent positive correlation between the sum of selenium and NAC concentrations and mean sperm concentration, motility, and normal morphology (17).

*Folic Acid*: In a double-blind, placebo-controlled interventional study, subfertile men receiving combined therapy of folic acid and zinc showed a statistically significant 74% increase in total normal sperm concentration after 26 weeks of treatment (18).

*Selenium*: A randomized placebo-controlled clinical trial involving 468 infertile men with idiopathic oligoasthenozoospermia demonstrated significant improvements in all semen parameters with selenium (200 mg) alone, NAC (600 mg) alone, or a combination of both supplements compared to placebo (17). The combination of selenium with vitamin E resulted in increased sperm motility (24). However, a study with normozoospermic men using selenium (300 mg) daily for 48 weeks did not show a significant influence on semen parameters (29).

Zinc: In a prospective trial with asthenozoospermic men, zinc supplementation for 3 months led to a significant improvement in sperm concentration, progressive motility, fertilizing capacity, and a reduction in the incidence of anti-sperm antibodies (19). Oral zinc supplementation restored seminal catalase-like activity and improved sperm concentration and progressive motility in asthenozoospermic men (49).

*Lycopene*: In a study involving 30 men with idiopathic oligoasthenozoospermia, treatment with 2 mg lycopene twice daily for 3 months resulted in statistically significant improvements in sperm concentration and motility in 66% and 53% of patients, respectively (33). A similar dose of lycopene was used in the treatment of 50 patients

with idiopathic oligoasthenozoospermia, and after a 1year follow-up, sperm concentration, motility, and morphology improved in 70%, 54%, and 38% of patients, respectively (35).

Antioxidant influence on advanced sperm function tests The conventional semen analysis has faced criticism for its limited ability to predict fertility accurately. While it offers valuable information on sperm production, accessory organ secretions, ejaculation, and emission, it falls short in predicting fertility (50). It does not provide insights into the functional potential of sperm to successfully fertilize an ovum or undergo the necessary maturation processes for fertilization. To address this limitation, advanced tests of sperm function were developed to enhance the predictive power of semen studies. Among these advanced tests, SDF and OS measures have been the most widely studied. Recent research has expanded our understanding of the implications of SDF on male fertility (8). Human sperm DNA, mostly bound to protamine, forms a condensed chromatin that is easily transportable through the sperm head and more resistant to damage during transit through the reproductive tracts (51). However, SDF can occur due to errors in chromatin packaging during spermatogenesis or exposure to seminal OS during epididymal transit (52). Both in vitro and in vivo studies confirm that elevated SDF can negatively impact fertility at various stages, including fertilization, early embryo development, implantation, and pregnancy (53,54). Therefore, addressing OS appears to be a justifiable approach to minimize SDF incidence in semen samples. Several studies have investigated the impact of dietary antioxidant supplementation on sperm DNA integrity (22, 25, 36, 55). While these studies generally assessed small-sized samples and had short treatment durations, they consistently reported a positive effect on SDF measures. For instance, Greco et al. (20) found a significant reduction in SDF percentage (p < 0.001) in patients with unexplained infertility and elevated SDF levels treated with vitamin C and vitamin E. Another study reported a 19% decrease in SDF (p < 0.001) with a combination of antioxidants containing zinc and selenium (36). Abad et al. (29) examined the effects of oral antioxidant therapy on SDF dynamics, revealing significant reductions at each experimental time-point (p < 0.05). Studies also explored antioxidant therapy in patients with high SDF due to varicocele. In one study, a combined antioxidant regimen led to a significant decrease in SDF levels (22.1%, p = 0.02) and an increase in sperm concentration (p = 0.04) (38). Assessing seminal OS levels has become integral in evaluating infertile men, considering its utility in various clinical scenarios (56). However, routine clinical use is hindered by factors such as test availability, complexity, cost-effectiveness, and a lack of universally accepted analysis methods. Various assays, classified as direct (e.g., chemiluminescence and flow cytometry assays) and indirect (e.g., myeloperoxidase test, lipid peroxidation levels), are available to measure OS. Each type has its advantages and disadvantages, with direct assays providing accurate measures but being expensive and requiring expertise, while indirect assays are simpler and more cost-effective but

assess an end state influenced by various unknown pathological processes (57-59). Numerous studies investigating the impact of antioxidant therapy on male fertility have evaluated its effects on OS as a key outcome measure. For instance, a 6-month regimen of vitamin E (300 mg daily) significantly reduced lipid peroxidation in semen samples from 110 asthenozoospermic men (23). Similarly, Comhaire et al. (24) reported a significant decrease in seminal ROS levels with a combination of 180 mg vitamin E and 30 mg  $\beta$ -carotene in 27 infertile men. Omu et al. (17) examined the effectiveness of daily supplementation with vitamin E (20 mg), vitamin C (10 mg), and zinc (400 mg) over 3 months in 45 asthenozoospermic men, observing a twofold reduction in malondialdehyde (an indicator of lipid peroxidation) (p < 0.01), a significant decrease in pro-apoptosis markers (p < 0.05), and a substantial increase in total antioxidant capacity (P < 0.01). Another study revealed that vitamin E (400 mg) and selenium (225 mg) intake for 3 months led to significant reductions in malondialdehyde levels, coupled with improvements in sperm motility and viability (24). Furthermore, Oeda et al. (60) observed a direct dose- and time-dependent reduction in seminal ROS when semen samples were incubated with N-acetylcysteine (NAC), suggesting the potential usefulness of NAC in reducing OS. In a randomized placebo-controlled study involving 120 patients with idiopathic infertility, those receiving 600 mg of NAC daily showed significant improvements in sperm motility compared to the placebo group (61). Additionally, Gharagozloo and Aitken (62) conducted a systematic review of 20 trials, indicating a significant reduction in OS or sperm DNA damage after antioxidant treatment in 19 of them.

## CONCLUSIONS

Numerous research has investigated the impact of antioxidant therapy on male fertility, demonstrating its potential in reversing OS-induced sperm dysfunction. Commonly used compounds include vitamin E, vitamin C, carnitines, zinc, selenium, NAC, CoQ10, folic acid, and lycopene. However, the identification of an ideal antioxidant treatment method is hindered by study design heterogeneity and the unknown normal physiological level of the fine redox balance. Further studies are necessary to determine the optimal and safe antioxidant preparation for managing male infertility.

## REFERENCES

1. Ahmad Majzoub and Ashok Agarwal. Systematic review of antioxidant types and doses in male infertility: Benefits on semen parameters, advanced sperm function, assisted reproduction and live-birth rate. Arab J Urol 2018; 16:113-124.

2. Agarwal A, Mulgund A, Hamada A, Chyatte MR. A unique view on male infertility around the globe. Reprod Biol Endocrinol 2015; 13:37.

3. Halliwell B. Free radicals and vascular disease: how much do we know? BMJ 1993; 307:885-886.

4. Brooker RJ. Genetics: analysis and principles. 4th ed. Ohio, USA: McGraw-Hill Higher Education; 2011.

5. Aitken RJ, Clarkson JS, Fishel S. Generation of reactive oxygen species, lipid peroxidation, and human sperm function. Biol Reprod 1989; 41:183-197.

6. Sies H. Strategies of antioxidant defence. Eur J Biochem 1993; 215:213-219.

7. Agarwal A, Majzoub A, Esteves SC, et al. Clinical utility of sperm DNA fragmentation testing: practice recommendations based on clinical scenarios. Transl Androl Urol 2016; 5:935-950.

8. Agarwal A, Sharma RK, Nallella KP, et al. Reactive oxygen species as an independent marker of male factor infertility. Fertil Steril 2006; 86:878-885.

9. Agarwal A, Saleh RA, Bedaiwy MA. Role of reactive oxygen species in the pathophysiology of human reproduction. Fertil Steril 2003; 79:829-843.

10. Gharagozloo P, Gutierrez-Adan A, Champroux A, et al. A novel antioxidant formulation designed to treat male infertility associated with oxidative stress: promising preclinical evidence from animal models. Hum Reprod 2016; 31:252-256.

11. ElSheikh MG, Hosny MB, Elshenoufy A, et al. Combination of vitamin E and clomiphene citrate in treating patients with idiopathic oligoasthenozoospermia: a prospective, randomized trial. Andrology 2015; 3:864-867.

12. Paradiso Galatioto G, Gravina GL, Angelozzi G, et al. May antioxidant therapy improve sperm parameters of men with persistent oligospermia after retrograde embolization for varicocele? World J Urol 2008; 26:97-102.

13. Peivandi S, Karimpour A, Moslemizadeh N. Effects of L-carnitine on infertile men's spermogram; a randomized clinical trial. J Reprod Infertil 2010; 10:245-251.

14. Safarinejad MR. Efficacy of coenzyme Q10 on semen parameters, sperm function and reproductive hormones in infertile men. J Urol 2009; 182:237-248.

15. Safarinejad MR, Safarinejad S. Efficacy of selenium and/or N-acetyl-cysteine for improving semen parameters in infertile men: a double-blind, placebo controlled, randomized study. J Urol 2009; 181:741-751.

16. Wong WY, Merkus HM, Thomas CM, et al. Effects of folic acid and zinc sulfate on male factor subfertility: a double-blind, randomized, placebo-controlled trial. Fertil Steril 2002; 77:491-498.

17. Omu AE, Al-Azemi MK, Kehinde EO, et al. Indications of the mechanisms involved in improved sperm parameters by zinc therapy. Med Princ Pract 2008; 17:108-116.

18. Lenzi A, Sgrò P, Salacone P, et al. A placebo-controlled doubleblind randomized trial of the use of combined l-carnitine and lacetyl-carnitine treatment in men with asthenozoospermia. Fertil Steril 2004; 81:1578-1584.

19. Ciftci H, Verit A, Savas M, et al. Effects of Nacetylcysteine on semen parameters and oxidative/antioxidant status. Urology 2009; 74:73-76.

20. Greco E, Iacobelli M, Rienzi L, et al. Reduction of the incidence of sperm DNA fragmentation by oral antioxidant treatment. J Androl 2005; 26:349-353.

21. Suleiman SA, Ali ME, Zaki ZM, et al. Lipid peroxidation and human sperm motility: protective role of vitamin E. J Androl 1996; 17:530-537.

22. Keskes-Ammar L, Feki-Chakroun N, Rebai T, et al. Sperm oxidative stress and the effect of an oral vitamin E and selenium supplement on semen quality in infertile men. Arch Androl 2003; 49:83-94. 23. Tremellen K, Miari G, Froiland D, Thompson J. A randomised control trial examining the effect of an antioxidant (Menevit) on pregnancy outcome during IVF-ICSI treatment. Aust N Z J Obstet Gynaecol 2007; 47:216-221.

24. Comhaire FH, Christophe AB, Zalata AA, et al. The effects of combined conventional treatment, oral antioxidants and essential fatty acids on sperm biology in subfertile men. Prostaglandins Leukot Essent Fatty Acids 2000; 63:159-165.

25. Gupta NP, Kumar R. Lycopene therapy in idiopathic male infertility - a preliminary report. Int Urol Nephrol 2002; 34:369-372.

26. Moslemi MK, Tavanbakhsh S. Selenium-vitamin E supplementation in infertile men: effects on semen parameters and pregnancy rate. Int J Gen Med 2011; 4:99-104.

27. Mohanty NK, Kumar S, Jha AK, Arora RP. Management of idiopathic oligoasthenospermia with lycopene. Indian J Urol 2001; 18:57-61.

28. Ménézo YJ, Hazout A, Panteix G, et al. Antioxidants to reduce sperm DNA fragmentation: an unexpected adverse effect. Reprod Biomed Online 2007; 14:418-421.

29. Abad C, Amengual MJ, Gosálvez J, et al. Effects of oral antioxidant treatment upon the dynamics of human sperm DNA fragmentation and subpopulations of sperm with highly degraded DNA. Andrologia 2013; 45:211-216.

30. Gual-Frau J, Abad C, Amengual MJ, et al. Oral antioxidant treatment partly improves integrity of human sperm DNA in infertile grade I varicocele patients. Hum Fertil (Camb) 2015; 18:225-229.

31. Singh F, Charles AL, Schlagowski AI, et al. Reductive stress impairs myoblasts mitochondrial function and triggers mitochondrial hormesis. BBA 2015; 1853:1574-1585.

32. Mentor S, Fisher D. Aggressive antioxidant reductive stress impairs brain endothelial cell angiogenesis and blood brain barrier function. Curr Neurovasc Res 2017; 14:71-81.

33. Lamosova D, Jurani M, Greksak M, et al. Effect of Rooibos tea (Aspalathus linearis) on chick skeletal muscle cell growth in culture. Comp Biochem Physiol C: Pharmacol Toxicol Endocrinol 1997; 116:39-45.

34. Omu AE, Fatinikun T, Mannazhath N, Abraham S. Significance of simultaneous determination of serum and seminal plasma alphatocopherol and retinol in infertile men by high-performance liquid chromatography. Andrologia 1999; 31:347-354.

35. Jacob RA, Pianalto FS, Agee RE. Cellular ascorbate depletion in healthy men. J Nutr 1992; 122:1111-1118.

36. Banihani S, Agarwal A, Sharma R, Bayachou M. Cryoprotective effect of L-carnitine on motility, vitality and DNA oxidation of human spermatozoa. Andrologia 2014; 46:637-641.

37. Lewin A, Lavon H. The effect of coenzyme Q10 on sperm motility and function. Mol Aspects Med 1997; 18(Suppl.):S213-219.

38. Erkkila" K, Hirvonen V, Wuokko E, et al. N-acetyl-L-cysteine inhibits apoptosis in human male germ cells in vitro. J Clin Endocrinol Metab 1998; 83:2523-2531.

39. Ursini F, Heim S, Kiess M, et al. Dual function of the selenoprotein PHGPx during sperm maturation. Science 1999; 285:1393-1396.

40. Hambidge KM, Krebs NF. Zinc deficiency: a special challenge. J Nutr 2007; 137:1101-1105.

41. Joshi R, Adhikari S, Patro BS, et al. Free radical scavenging

behavior of folic acid: evidence for possible antioxidant activity. Free Radic Biol Med 2001; 30:1390-399.

42. Agarwal A, Sekhon LH. Oxidative stress and antioxidants for idiopathic oligoasthenoteratospermia: Is it justified? Indian J Urol 2011; 27:74-85.

43. Cooper TG, Noonan E, von Eckardstein S, et al. World Health Organization reference values for human semen characteristics. Hum Reprod Update 2010; 16:231-245.

44. Dawson EB, Harris WA, Teter MC, Powell LC. Effect of ascorbic acid supplementation on the sperm quality of smokers. Fertil Steril 1992; 58:1034-1039.

45. Cyrus A, Kabir A, Goodarzi D, Moghimi M. The effect of adjuvant vitamin C after varicocele surgery on sperm quality and quantity in infertile men: a double blind placebo controlled clinical trial. Int Braz J Urol 2015; 41:230-238.

46. Balercia G, Regoli F, Armeni T, et al. Placebo-controlled doubleblind randomized trial on the use of L-carnitine, L-acetylcarnitine, or combined L-carnitine and L-acetylcarnitine in men with idiopathic asthenozoospermia. Fertil Steril 2005; 84:662-671.

47. Nadjarzadeh A, Shidfar F, Amirjannati N, et al. Effect of Coenzyme Q10 supplementation on antioxidant enzymes activity and oxidative stress of seminal plasma: a double-blind randomised clinical trial. Andrologia 2014; 46:177-183.

48. Lafuente R, González-Comadrán M, Solà I, et al. Coenzyme Q10 and male infertility: a meta-analysis. J Assist Reprod Genet 2013; 30:1147-1156.

49. Hadwa MH, Almashhedy LA, Alsalman AR. Oral zinc supplementation restores superoxide radical scavengers to normal levels in spermatozoa of Iraqi asthenospermic patients. Int J Vitam Nutr Res 2015; 85:165-173.

50. Esteves SC. Clinical relevance of routine semen analysis and controversies surrounding the 2010 World Health Organization criteria for semen examination. Int Braz J Urol 2014; 40:443-453.

51. Erenpreiss J, Spano M, Erenpreisa J, et al. Sperm chromatin structure and male fertility: biological and clinical aspects. Asian J Androl 2006; 8:11-29.

52. Shamsi MB, Kumar R, Dada R. Evaluation of nuclear DNA damage in human spermatozoa in men opting for assisted reproduction. Indian J Med Res 2008; 127:115-123.

53. Sharma RK, Said T, Agarwal A. Sperm DNA damage and its clinical relevance in assessing reproductive outcome. Asian J Androl 2004; 6:139-148.

54. Saleh RA, Agarwal A, Sharma RK, et al. Evaluation of nuclear DNA damage in spermatozoa from infertile men with varicocele. Fertil Steril 2003; 80:1431-1436.

55. Saleh RA, Agarwal A, Nada EA, et al. Negative effects of increased sperm DNA damage in relation to seminal oxidative stress in men with idiopathic and male factor infertility. Fertil Steril 2003; 79(Suppl. 3):1597-1605.

56. Agarwal A, Cho CL, Esteves SC. Should we evaluate and treat sperm DNA fragmentation? Curr Opin Obstet Gynecol 2016; 28:164-171.

57. Gil-Villa AM, Cardona-Maya W, Agarwal A, et al. Role of male factor in early recurrent embryo loss: do antioxidants have any effect? Fertil Steril 2009; 92:565-571.

58. Agarwal A, Makker K, Sharma R. Clinical relevance of oxidative stress in male factor infertility: an update. Am J Reprod Immunol 2008; 59:2-11.

59. Agarwal A, Tvrda E, Sharma R. Relationship amongst terato-

Archivio Italiano di Urologia e Andrologia 2024; 96(2):12323

zoospermia, seminal oxidative stress and male infertility. Reprod Biol Endocrinol 2014; 12:45.

60. Oeda T, Henkel R, Ohmori H, Schill WB. Scavenging effect of Nacetyl-L-cysteine against reactive oxygen species in human semen: a possible therapeutic modality for male factor infertility? Andrologia 1997; 29:125-131.

61. Benatta M, Kettache R, Buchholz N, Trinchieri A. The impact of nutrition and lifestyle on male fertility. Arch Ital Urol Androl. 2020; 92.121-131.

62. Gharagozloo P, Aitken RJ. The role of sperm oxidative stress in male infertility and the significance of oral antioxidant therapy. Hum Reprod 2011; 26:1628-1640.

#### Correspondence

Tamer A. Abouelgreed, MD (Corresponding Author) dr\_tamer\_ali@yahoo.com; tamerali.8@azhar.edu.eg Department of Urology, Al-Azhar University, Cairo, Egypt & Gulf medical university, Ajman, UAE

Mohamed A. Amer, MD amerrom@yahoo.com Hassan Mamdouh, MD hsdermaclinic@yahoo.com Hany Aboelwafa, MD dr\_hanyos138@yahoo.com Sameh F. Fahmy samehmohamed74@azhar.edu.eg Omar A. Omar, MD omarabdelhady.236@azhar.edu.eg Mohammed Abdelshakour, MD Dr.mohammed\_121@yahoo.com Mohammad Elesawy, MD elesawy288@gmail.com Mohamed Sonbol, MD bosombol1185@gmail.com Ahmed N. Maawad, MD ah.nabil70@gmail.com Osama K. Elsayed Osamaandroderma@gmail.com Department of Dermatology & Andrology Al-Azhar University, Cairo, Egypt

Ahmed F. El-Sherbiny, MD - Ahmed\_derma@yahoo.com Department of Andrology, International Islamic Center for Population Studies and Research, Al-Azhar University, Cairo, Egypt

Conflict of interest: The authors declare no potential conflict of interest.